

A review of asteroid biology in the context of sea star wasting: possible causes and consequences

RH: The biology of sea star wasting

5

Accepted version, pre proofs, in press at The Biological Bulletin

Authors

Nathalie Oulhen¹, Maria Byrne^{2,3}, Paige Duffin⁴, Marta Gomez-Chiarri⁵, Ian Hewson⁶, Jason Hodin⁷, Brenda Konar⁸, Erin K. Lipp⁹, Benjamin G. Miner¹⁰, Alisa L. Newton¹¹, Lauren M. Schiebelhut¹², Roxanna Smolowitz¹³, Sarah J. Wahltinez¹⁴, Gary M. Wessel¹, Thierry M. Work¹⁵, Hossam A. Zaki¹, John P. Wares*^{4,16}

* Corresponding author: jpwares@uga.edu

Affiliations

15 ¹Department of Molecular and Cell Biology and Biochemistry, Brown University, Providence, Rhode Island, USA

²School of Medical Sciences, The University of Sydney, Sydney, New South Wales, Australia

20 ³School of Life and Environmental Sciences, The University of Sydney, Sydney, New South Wales, Australia

⁴Department of Genetics, University of Georgia, Athens, Georgia, USA

25 ⁵Department of Fisheries, Animal and Veterinary Science, University of Rhode Island, Kingston, Rhode Island, USA

⁶Department of Microbiology, Cornell University, Ithaca, New York, USA

⁷Friday Harbor Labs, University of Washington, Friday Harbor, WA, USA

30 ⁸College of Fisheries and Ocean Sciences, University of Alaska Fairbanks, Fairbanks, Alaska, USA

⁹Department of Environmental Health Science, University of Georgia, Athens, Georgia, USA

¹⁰Department of Biology, Western Washington University, Bellingham, Washington, USA

35 ¹¹Disney's Animals Science and Environment, Orlando, FL, USA

¹²Department of Life and Environmental Sciences, University of California, Merced, CA, USA

40 ¹³Department of Biology and Marine Biology, Roger Williams University, Bristol, Rhode Island, USA

¹⁴Department of Comparative, Diagnostic, and Population Medicine, College of Veterinary Medicine, University of Florida, Gainesville, FL, USA

45 ¹⁵U.S. Geological Survey, National Wildlife Health Center, Honolulu Field Station, Honolulu, HI 96850, USA.

¹⁶Odum School of Ecology, University of Georgia, Athens, Georgia, USA.

Abstract

Sea star wasting – marked in a variety of sea star species as varying degrees of skin lesions followed by disintegration – recently caused one of the largest marine die-offs ever recorded
55 on the west coast of North America, killing billions of sea stars. Despite the important ramifications this mortality had for coastal benthic ecosystems, such as increased abundance of prey, little is known about the causes of the disease or mechanisms of its progression. Though there have been studies indicating a range of causal mechanisms, including viruses and environmental effects, the broad spatial and depth range of affected populations leaves
60 many questions remaining about either infectious or non-infectious mechanisms. Wasting appears to start with degradation of mutable connective tissue in the body wall, leading to disintegration of the epidermis. Here, we briefly review basic sea star biology in the context of the sea star wasting syndrome and present our current knowledge and hypotheses related to the symptoms, the microbiome, the viruses, and associated environmental stressors. We
65 also highlight throughout the article knowledge gaps and the data needed to better understand sea star wasting mechanistically, its causes, and potential management.

Introduction

In the past decade, sea stars (Echinodermata: Class Asteroidea, “asteroids”) have been
75 confronted with a pandemic that has gained increasing attention because of the severity,
taxonomic breadth, and geographic extent of the disease. Previously referred to as sea star
wasting disease (SSWD), sea star wasting syndrome (SSWS), asteroid idiopathic wasting
syndrome (AIWS), or just sea star wasting (SSW), the nomenclature points to a suite of signs
80 observed across a broad range of species that, in the most severe cases, result in death
following disintegration of the star (Hewson *et al.*, 2018). Although species-specific
variations exist, wasting sea stars generally exhibit modified behavior, such as lethargy and
disinterest in nearby prey, pronounced body wall lesions, and arm autotomy (Figure 1;
Eisenlord *et al.*, 2016, Kohl *et al.*, 2016, Menge *et al.*, 2016, Jaffe *et al.*, 2019).

85 While the primary SSW event that is often referred to in current discussion – emerging on the
Pacific coast of North America in 2013 – is the most extreme documented example, prior
asteroid wasting events matching this general description have been periodically observed,
with the first known reported over a century ago (Mead, 1898). Over the years, wasting has
been noted independently across many sea star species globally, often with regional
90 mortality, including the North American Pacific (Dungan *et al.*, 1982, Eckert *et al.*, 2000,
Konar *et al.*, 2019), and Atlantic coasts (Sieling, 1960, Tiffany, 1978, Menge, 1979, Bucci *et al.*,
2017), Mediterranean Sea (Stahli *et al.*, 2009), Atlantic coast of Europe (Thorpe and
Spencer, 2000), East China Sea (Suzuki *et al.*, 2012), Yellow Sea (China), and Port Philip
Bay, Australia (Hewson *et al.*, 2019). Importantly, most earlier events targeted only a few
95 species in an area. For example, in the summer of 1978, populations of *Heliaster kubiniji*
(sun star) suffered massive mortality in the Gulf of California rocky intertidal zone. The
outbreak reduced abundance of *H. kubiniji* from dense aggregations to near-zero values at
sites throughout the Gulf of California in just a few weeks (Dungan *et al.*, 1982); there was
similarly high intertidal mortality in the smaller high intertidal sea star *Othilia tenuispina*, but
100 this species rebounded in numbers quickly. At least four co-occurring species of subtidal sea
stars were reportedly unaffected. The event was coincident with exposure to warmer waters
in the intertidal (Dungan *et al.*, 1982). In contrast, the occurrence of SSW in the past decade

has profoundly affected more than 20 species of sea stars in the northeastern Pacific and other sites— from intertidal to deeper waters, from Baja California to northern Gulf of Alaska, and beyond (Dawson *et al.*, this issue). In this sense, the current prevalence of wasting is among the most geographically and taxonomically widespread diseases of invertebrates ever recorded.

Many asteroids are top level consumers with direct effects on the structure and function of benthic ecosystems (Menge *et al.*, 1994, Vicknair and Estes, 2012, Schultz *et al.*, 2016, Burt *et al.*, 2018). Indeed, the concept of keystone species was first developed from work on populations of the predatory *Pisaster ochraceus* in the Pacific Northwest (Paine, 1966). When present, *P. ochraceus* consumes the dominant encrusting invertebrate species, thus freeing up space for other sessile macroinvertebrates and algae (Feder, 1959, McClintock and Robnett, 1986, Robles *et al.*, 2009, Lafferty and Suchanek, 2016). Other asteroid species function similarly, sometimes at different size scales (Gravem and Morgan, 2017). In addition to direct predation, some asteroids have important indirect effects by altering prey behavior (Duggins, 1983, Rogers *et al.*, 2018, Kay *et al.*, 2019, Murie and Bourdeau, 2019). Sea stars also directly compete for marine resources with other high-level consumers (Traiger *et al.*, 2016), and can have substantial effects on aquaculture and restoration endeavors (Silina, 2008, Miyoshi *et al.*, 2018, Wilcox and Jeffs, 2019).

Along with the key ecosystem functions of many echinoderms, mass mortality events can generate cascading effects, ranging from temporary local disruptions to long-term ecological phase shifts (Lessios *et al.*, 1984, Burge *et al.*, 2014, Miner *et al.*, 2018). Mass mortality events affecting critical foundation species, ecosystem engineers, or keystone species have been coined ‘marine disease emergencies’ due to the detrimental, and sometimes irreversible, series of events that often succeeds them (Miner *et al.*, 2018). A textbook example of coincident marine disease events with profound ecological effects started with mass die-offs of the sea urchin *Diadema antillarum* that resulted in overgrowth of algae on coral reefs (Lessios *et al.*, 1984) and the subsequent mass mortality through a variety of causes of *Acropora* corals led to complete physical restructuring of Caribbean coral reefs (Hughes, 1994, Hughes *et al.*, 2003, Cramer *et al.*, 2020). Notably, all of these changes promote

135 increasing prevalence of microbial and algal nutrient cycles, a feature of many recent
climate-driven shifts leading to eutrophication of marine ecosystems (Petraitis and Dudgeon,
2020).

140 In this context, recent mass mortality events affecting asteroids threaten the health of coastal
ecosystems and additional understanding is needed to manage or mitigate them. As stated
above, the most recent SSW event has differed markedly from prior events in their
geographic and taxonomic extent. This epizootic killed many billions of asteroids across over
20 taxa (indeed, billions are estimated to have been lost in a single species alone; e.g.,
Pycnopodia helianthoides (Gravem *et al.*, 2021)), primarily along the Pacific coast of North
America where the coastal community is regularly monitored, and is considered to be among
145 the most geographically widespread disease events to sweep through marine wildlife in
documented history (Gudenkauf and Hewson, 2015, Eisenlord *et al.*, 2016, Hewson *et al.*,
2018). SSW has been forecast to reshape community structure throughout the region (Menge
et al., 2016, Gravem and Morgan, 2017) and due to its broad taxonomic effect, is considered
a risk for other coastal communities globally (Hewson *et al.*, 2019).

150 Unfortunately, there is a deficit of fundamental knowledge surrounding the etiology,
pathogenesis, and epidemiology of SSW, reflected in the sparse and discontinuous literature
on the subject. Thus, in an attempt to generate the framework for a cohesive narrative on
SSW for future reference, the aims of our review are to: (1) provide a relevant review of
155 normal sea star biology, anatomy, and physiology in the context of the common outward
signs of SSW in affected sea stars and what we have learned, (2) highlight our understanding
of the biotic factors associated with wasting including microbial and viral interactions, (3)
identify crucial gaps in our understanding of the symptoms and progression of SSW, and (4)
point to forthcoming research and point to forthcoming research designed to further our
160 understanding of SSW.

We begin by considering the basic biology of sea stars. Our goal in this first section is to
establish which anatomical systems may be initially or overtly affected by SSW, in a way
that informs not only the recent and unprecedented mass mortality event, but also prior

165 historical examples. Our paper then reviews each of these organismal systems for indicators
of cause or response to SSW.

170 **Asteroid biology as context for SSW**

In this section, we review some basic elements of sea star biology to introduce unique
aspects, traits, and physiology that come into play – across large phylogenetic breadth
(Schiebelhut *et al.*, this issue) – when these animals succumb to SSW. The ecological
175 consequences noted above are intertwined with both the density and distribution of sea stars
as well as the broadly defined traits that are affected by SSW. Sea stars are widely distributed
throughout the world’s oceans. They are stenohaline and occur from the deep sea to intertidal
zone, and from tropical to polar waters (Binyon, 1961, Gage and Tyler, 1991, Byrne *et al.*,
2017). Across this extensive range, local species richness is highly variable (Cintra-
180 Buenrostro *et al.*, 2005, Iken *et al.*, 2010), with diversity driven by many environmental and
ecological factors. Local diversity can encompass deep phylogenetic divergences, as often
dozens of genera and several families can be found within any ecoregion (*sensu* Spalding *et al.*,
2007) – this is key because observed effects of SSW affect such great diversity of sea star
species (see accompanying papers in this issue) with vastly different life histories and diets
185 (VanVeldhuizen and Oakes, 1981, McClintock *et al.*, 2008, Lawrence, 2015, Martinez *et al.*,
2017, Pratchett *et al.*, 2017).

The size, density, and distribution of sea star populations are highly variable through time
(Uthicke *et al.*, 2009), but they are not always closely monitored in the absence of clearly
190 identified economic consequences (Hewson *et al.*, 2019). Nevertheless, demographic variation in
sea star populations is associated with recognizable ecological responses such as overgrowth by
mussels in the absence of sea star predation (Paine, 1966, Gravem and Morgan, 2017, Kay *et al.*,
2019). Some distributional patterns are difficult to track because they comprise multiple cryptic
species. For instance, species of the *Acanthaster* complex are found throughout tropical waters
195 and can form high-density feeding fronts that devastate coral reefs (Pratchett *et al.*, 2017), but the

individual *Acanthaster* species and their distributions remain uncertain (Haszprunar *et al.*, 2017). Similar problems have been noted in the taxonomy – and thus the precision in evaluating disease effects – of other asteroid genera (Melroy and Cohen, 2021). What is important about these components of phylogenetic diversity and potential density-dependent effects is not yet clear, but
200 if we are to understand the apparent transmissibility of SSW (Hewson *et al.* 2014) then we must know how and whether the health of any sea star in an ecosystem is predictive of the health of other individuals, both conspecific and across the phylogeny of asteroids (Schiebelhut *et al.*, this issue). Whether SSW is primarily driven by infectious or non-infectious etiologies is still unclear – and disease etiology likely involves complex interactions – but the broad taxonomic effect
205 indicates the shared traits of sea stars could be a first target for clues into mechanisms of pathogenesis.

Body plan and life history

210 Echinoderms possess four main features that distinguish them from other metazoans: their calcareous endoskeleton composed of ossicles of varying sizes, a water vascular system, mutable collagenous tissues, and pentaradial body organization in adults (Hyman, 1955, Sloan, 1980, Jangoux, 1982, Blake, 1989, Chia and Koss, 1994, Byrne, 1999, Mooi and
215 David, 2000, McEdward and Miner, 2001, McEdward *et al.*, 2002, Byrne, 2013, Lawrence 2013, Mercier and Hamel, 2013, Martinez *et al.*, 2016). The present-day global asteroid fauna includes approximately 1,900 species in 37 families (Mah and Blake, 2012, Feuda and Smith, 2015, Byrne *et al.*, 2017, Linchangco Jr *et al.*, 2017).

220 Asteroid life histories have been studied across diverse taxa (Strathmann, 1985, Chia *et al.*, 1993, McEdward and Miner, 2001, McEdward *et al.*, 2002, Byrne, 2013), though most such studies are biased towards those taxa that are most available near prominent marine research facilities. Most species are dioecious free-spawners that release their gametes into the water column for fertilization, followed by development of a planktonic larva (Emlet *et al.*, 1987),
225 and then metamorphosis and settlement back to the sea floor. Some species deposit eggs that adhere to rocks on the shore and have benthic development (Chia, 1968, Byrne, 1995). Other

asteroids retain their eggs in brood chambers on or within the body cavity (Chia *et al.*, 1993, McEdward and Janies, 1993). Variation in reproductive strategies such as these may influence the capacity of a species to recover from mass mortality events. There does not appear to be a close link between larval dispersal and SSW (Schiebelhut *et al.*, this issue), and it is still unclear if early life stages are susceptible to – or could transmit - the disease.

Surface and skeleton

The breakdown of the body surface and its integrity is a feature of SSW, and so an understanding of these tissues is important for understanding the disease. The surface of a sea star is one of the more complicated types of external barriers in the Metazoa. The surface can be covered by abundant spines, or have no spines in species with a thick, soft epidermis (e.g., *Dermasterias imbricata*). The surface consists of an external columnar epithelium (containing secretory cells, supporting cells, sensory cells, and glandular cells). The epithelial cells are covered by a thin cuticle that is only visible microscopically (McKenzie and Grigolava, 1996). The body wall connective tissue has mutable properties (MCT - mutable connective tissue), is capable of reversible rapid changes of tensile strength from stiff to viscous, is under neural control, and is mediated by proteinaceous neurotransmitters (for review see Wilkie, 2002, Barbaglio *et al.*, 2013). Ossicles composed of magnesium calcite are embedded in the MCT and form the endoskeleton. This internal calcareous skeleton is considered a synapomorphy of the echinoderms (but see Cameron and Bishop, 2012).

In sea stars, MCT is found in the body wall dermis, spine joint ligaments, and in the epithelial cells of the tube feet (Motokawa, 1986, O'Neill, 1989, Santos *et al.*, 2005, Motokawa, 2011), and is composed of spindle-shaped collagen fibers in a proteoglycan/glycosaminoglycan matrix to which they are covalently bound (Motokawa, 1982, O'Neill, 1989, Trotter and Koob, 1989, Wilkie *et al.*, 1992, Erlinger *et al.*, 1993, Barbaglio *et al.*, 2012). Cells in the MCT include immune cells, nerve cells, and juxtaligamental cells that are unique to the MCT. The juxtaligamental cells are recognized by their possession of dark granules which are thought to have effector molecules involved in the nervous system innervation of MCT (Wilkie, 1979, 2002).

260 The coelomic cavity is a hemocoel lined by a layer of myoepithelial, secretory, and
supporting cells. This epithelium tissue also forms the papulae that project through the thick
body wall and into the overlying sea water. Papulae are covered both externally and
internally by the two epithelial layers noted above; however, the intervening connective
tissues between the epithelial layers are greatly thinned (Hyman, 1955, Cobb, 1978). Papulae
also can be retracted in recessed “pockets” with the use of muscles and cavities at the base of
265 the papula, or they can be extended quickly above the surface by relaxation of the muscles
and diversion of the fluid contents of the perivisceral coelom into the papular lumen. Papulae
function in oxygen exchange and waste disposal (Binyon, 1972, Cobb, 1978).

Pedicellariae are small pincer-like compound ossicles covered by epithelium and controlled
by muscles. Pedicellariae are found primarily on the aboral body surface, and function in
270 removal of small particulates or fouling organisms that settle on the body wall (Campbell
and Rainbow, 1977, Ruppert, 2004), defense (Jensen, 1966, Chia, 1969, VanVeldhuizen and
Oakes, 1981), aggression (Menge and Menge, 1974, Wobber, 1975), and prey capture
(Robilliard, 1971, Chia and Amerongen, 1975, Hendler and Franz, 1982, Dearborn, 1991,
Emson and Young, 1994, Lauerman, 1998).

275 On the ventral side of the sea star, the centrally-located mouth faces the benthos. Ambulacral
grooves bearing the tube feet radiate from the oral surface to the ends of each arm; the highly
distensible tube feet enable locomotion and prey capture. These may have terminal adhesive
suckers for attachment or be pointed in species that burrow in soft sediments (Paine, 1926,
Smith, 1937). Paired ambulacral ossicles (skeletal elements) form an arch-like arcade
280 spanning the entire arm and overlaying the water vascular canal and tube feet that project
into the ambulacral groove.

Nervous system

285 The nervous system organization reflects the pentaradial symmetry and is composed of a
circumoral nerve ring from which the radial nerve cords originate that run along the arms just
internal to the ambulacral grooves. At the end of each arm, the radial nerve cord connects

with the ocelli of the eye spot, which is a phototactic sensory organ. The nervous system is divided into ectoneural and hyponeural systems which are separated from one another by a thin connective tissue layer (Hyman, 1955, Cobb, 1987). The ectoneural system is mainly associated with sensory and motor components. The hyponeural tissue is considered to be exclusively motor and comprises the thinner inner layer of the radial nerve cords (Cobb, 1987). Perception of physical and chemical change in the environment influences hormonal and immune responses in echinoderms (Hamel *et al.*, 2021). Autotomy itself is a component sign of SSW; it is a complex behavioral response and can be induced by a number of diverse stressors (Alupay, 2013, Mladenov *et al.*, 1989; Byrne *et al.*, 2019). For this reason, much research has focused on how autotomy and nerve regeneration interplay, but the details remain elusive (Byrne, 2020).

300 ***Digestive system***

Sea stars have a complete digestive system. The mouth located toward the substrate is surrounded and controlled by a peristomial membrane. The mouth is connected to a short esophagus that leads to the cardiac stomach (Anderson, 1954, 1959). Sea stars can often evert their cardiac stomach to catch and digest their prey and retract the stomach afterward (Semmens *et al.*, 2013, Tinoco *et al.*, 2018). The cardiac stomach is separated from the pyloric stomach by a slight constriction and the pyloric stomach reaches the anus via a short intestine. The pyloric stomach also connects via a pyloric duct to a pair of pyloric caeca in each arm. These pyloric caeca, which consist of complex, ramified evaginations, have been described as the most important parts of the gut (both in volume and function) in asteroids (Binyon, 1972). Pyloric caeca provide the majority of digestive enzymes and also absorb and store nutrients (Ferguson, 1969). Their structures have been well described in *Asterias forbesi* and *Marthasterias glacialis* (Anderson, 1953, Martinez *et al.*, 1989). The connection between the digestive system and SSW is unclear. Sunflower stars (*Pycnopodia helianthoides*) in captivity that are fed a regular diet will only seem to refuse food when they are in poor condition – such as suffering from a wound – or otherwise stressed, such as shortly after collection. However, stars of this species have been observed to feed even when exhibiting early signs of ultimately fatal cases of SSW (J.H., unpublished)

320 *Coelomic system*

As is typical of echinoderms, asteroids have a hydraulic system known as the water vascular system, used for locomotion as well as particle transportation. This system consists of a central ring canal that connects with the radial canals which in turn connect with the tube feet. In asteroids, the stone canal connects the water vascular system with environmental water at the sieve plate called the madreporite (Hyman, 1955, Ferguson and Walker, 1991). Tube feet are connected to the radial canals through ampullae that can contract or relax to move water in each tube foot.

330 Within the body and surrounding all organs, including the water vascular system components, is the perivisceral coelom. The concentrations of electrolytes (Binyon, 1962, Stickle and Denoux, 1976, Held and Harley, 2009, Wahltinez *et al.*, 2019), amino acids, and ammonium (Diehl and Lawrence, 1984) in coelomic fluid are not substantially different from those in seawater (Binyon, 1962). Tube foot activity depends on the water vascular system and seems to be maintained by an active transport of potassium leading to an osmotic influx of water directly through their walls (Prusch and Whoriskey, 1976, Ferguson, 1990). While sea stars are osmoconformers, they are partially able to regulate coelomic fluid volume (Pearse, 1967, Ellington and Lawrence, 1974) though the mechanism remains unknown.

340 The hyponeural coelom runs parallel to the water vascular system from the madreporic ampulla to the lateral canals. The hyponeural coelom surrounds the vertical axial hemal canal, the hyponeural hemal ring, and the radial and lateral hemal vessels. The hyponeural coelom also forms the genital coelom that surrounds and connects the genital hemal ring and the gonads. The hyponeural canal and the stone canal both connect to the madreporic ampulla, and sea water is thought to supply both systems (Hyman, 1955, Ferguson and Walker, 1991). The “flattening” that is associated with SSW (see below) likely involves coelomic processes.

350 *The echinoderm immune system*

An important milestone in the identification of an immune system in biological organisms was reached in sea stars over a century ago. Eli Metchnikoff inserted a rose thorn into a sea star larva and to his surprise, saw cells migrate to this thorn and eventually encompass the intruder (see e.g., (Gordon, 2016b)). This was a novel characterization of the process of phagocytosis in an immune defense and helped earn Metchnikoff the Nobel Prize in 1908. Since then, our understanding of the immune system of this and other echinoderms has increased enormously; later sections will detail what we have learned because of SSW itself.

360 As with many invertebrates, host defenses comprise behavioral and cellular changes in response to non-self recognition, an intact epidermis with functional pedicellariae, and immune cells. Asteroids only have an innate system of immune defense. That is, cells and molecules are present in the organism that can recognize an invader as non-self and minimize infectivity by encapsulating, consuming through phagocytosis, or killing it (Smith *et al.*, 365 2010). Evidence for an adaptive system of immune memory is lacking. Limited analyses of available echinoderm genomes show a lack of elements known to adapt to pathogen exposure, e.g., (Sea Urchin Genome Sequencing *et al.*, 2006, Smith *et al.*, 2010, Hall *et al.*, 2017).

370 Much of the early work on immune function in sea stars was accomplished through grafting experiments. Grafts of a piece of tissue from one sea star to a different conspecific or heterospecific individual results in the host animal killing cells of the donor tissue at the border, and eventually releasing the graft (Hildemann and Dix, 1972, Karp and Hildemann, 1976). These similar responses demonstrate that individuals within a species can recognize 375 one another as non-self, and can do this in as robust a manner as seen in animals with adaptive immune mechanisms (Silva, 2000, Furukawa *et al.*, 2009).

Subsequent work on sea star immunity has focused on immune cells in the coelomic fluid and tissues, referred to as coelomocytes (Smith *et al.*, 2010). Asteroids have several types of 380 coelomocytes including phagocytic amoebocytes, phagocytes, amoebocytes, spherule cells,

vibratile cells, haemocytes, crystal cells, and progenitor cells (Smith, 1981). Each of these circulating cells appear to have unique characteristics and functions, in combination similar to the innate immune system functions of vertebrates, including clot formation, phagocytosis, encapsulation, clearance of bacteria or other foreign materials, and even oxygen transport
385 (Pinsino and Matranga, 2015, Smith *et al.*, 2018).

Innate immune systems rely on recognition of evolutionarily-conserved structures on pathogens, also known as pathogen-associated molecular patterns (PAMPs), which act through a limited number of pattern recognition receptors (PRRs), notably the family of Toll-like receptors (TLRs). As in other invertebrates, analyses of the sea urchin genome revealed a
390 profound complexity of innate immune recognition receptors, regulators, and effectors (Hibino *et al.*, 2006, Rast *et al.*, 2006, Smith *et al.*, 2018), generated perhaps by combinations of gene recombination, gene duplication, and selection for sequence diversification in response to high rates of change in their pathogens. Generally, these gene regions have been
395 identified in asteroid genomes as well (Rast *et al.*, 2006, Ruiz-Ramos *et al.*, 2020). While our insights are still limited to a few species that have been well-characterized, echinoderms have more than 10 times the number of gene copies in these families than found in vertebrates, providing a diverse innate immune complexity.

400 Like other ectotherms, environmental factors such as temperature, salinity, heavy metals, and UV light (Pinsino and Matranga, 2015) can affect the immune response of sea stars. Recent work has shown that the interaction of an echinoderm with stressors such as perceived predation rapidly influence the hormonal regulation of coelomocytes and other components of the innate immune system (Hamel *et al.*, 2021), and nutrient deprivation can drive
405 wasting-like responses in some sea stars (VanVolkom *et al.*, 2021). Thus, interpreting experimental data on immune and disease responses in echinoderms in the context of SSW would need to account and control for these environmental variables.

A challenge in our understanding of disease in echinoderms is the lack of basic knowledge of
410 cell physiology and mechanisms of host cell responses to insults. Moreover, there is a lack of

functional evidence to understand host cell gene expression and its role in cell death and genesis of lesions, all rich avenues for future investigative efforts.

Virology and Microbiology of grossly normal asteroids

415

Early microscopic study of apparently healthy asteroids found an abundance of subcuticular bacteria of varying morphologies (Holland and Nealson, 1978, Kelly *et al.*, 1995, Foret and Lawrence 2001). However, these studies highlighted the large abundance of spiral-shaped bacteria (which were distinctive for Spirochaetes) beneath the outer epidermis and embedded within body wall tissues (Holland and Nealson, 1978, Kelly *et al.*, 1995, Foret and Lawrence 420 2001). Cultivation-based surveys, which are highly biased towards taxa that grow on surfaces and in nutrient-rich conditions, reflected mostly copiotrophic taxa (i.e. microorganisms capable of rapid assimilation of compounds and subsequent rapid cell division) (e.g., Narita *et al.*, 1987, Choi *et al.*, 2003, Beleneva and Zhukova, 2009, Rivera-Posada *et al.*, 2011b, 425 Luo *et al.*, 2013, Hewson *et al.*, 2018), Table A1). However, most bacterial taxa in marine habitats cannot yet be cultivated; as a consequence, less-biased molecular barcoding approaches are typically used to examine the structure of microbial communities inhabiting marine habitats.

430 Despite the known cultivation biases, many commonly reported orders, families, and genera of bacteria in culture collections (Table A1) are also common constituents of cultivation-independent surveys (Nakagawa *et al.*, 2017, Hewson *et al.*, 2018, Hoj *et al.*, 2018, Jackson *et al.*, 2018, Lloyd and Pespeni, 2018, Nunez-Pons *et al.*, 2018; Table A2). These taxonomic groupings represent microorganisms that are typically found in organic nutrient-rich 435 environments in marine habitats.

By contrast, other aspects of asteroid-associated microdiversity (e.g., fungi and protists) have not been extensively documented. Nunez-Pons *et al.* (2018) examined fungal communities associated with grossly normal *Odontaster validus* and found that communities were dominated by asco- and basidiomycetes (notably, Saccharomycetes, Eurotiomycetes, 440 Dothideomycetes, and Agaricomycetes). The parasitic ciliate *Orchitophrya stellarum* is

known to infect testes of males and castrate them (Cepede, 1907, Peters, 1992, Lawrence, 2015) in a range of species (Vevers, 1951, Leighton *et al.*, 1991, Byrne *et al.*, 1997, Byrne *et al.*, 1998, Stickle *et al.*, 2001, Stickle and Kozloff, 2008, Sunday *et al.*, 2008). While *Orchitophrya* does not cause gross disease signs (Cepede, 1907, Bang, 1982), it may lead to mortality, especially for males (Leighton *et al.*, 1991, Claereboudt and Bouland, 1994) and reduced gamete abundance or fertility. Otherwise asymptomatic individuals may also be infected by an unidentified apicomplexan parasite reported in *Asterias amurensis* (Goggin and Bouland, 1997). Asteroids may host labyrinthulids (Fiorito *et al.*, 2016) and mesomycetozoa (Kerk *et al.*, 1995, Ragan *et al.*, 1996, Glockling *et al.*, 2013, Paps *et al.*, 2013, Hewson and Sewell, 2021). The functions of these less well-studied organisms in sea star health and disease are unclear, and they raise questions of whether such diversity is involved in SSW transmission or symptomatic progression.

Viruses inhabiting tissues of grossly normal sea stars are diverse and include both DNA and RNA viruses (Hewson *et al.*, 2014, Jackson *et al.*, 2016, Hewson *et al.*, 2018, Jackson *et al.*, 2020a). While much early attention in the SSW outbreak focused on the sea star associated densovirus (SSaDV, renamed Asteroid ambidensovirus-1 by the International Committee on Taxonomy of Viruses; Hewson *et al.*, 2014), this viral genotype was one of many inhabiting clinically normal asteroids, and appears to be a common constituent across grossly normal echinoderms worldwide (Gudenkauf *et al.*, 2014, Hewson *et al.*, 2018, Jackson *et al.*, 2020a, Jackson *et al.*, 2020b, Hewson and Sewell, 2021). Recent work has demonstrated that these taxa may develop persistent infections (Jackson *et al.*, 2020a). Asymptomatic individuals also bear picornaviruses, dicistroviruses (Hewson *et al.*, 2018, Hewson *et al.*, 2020a), circular rep-encoding single-stranded DNA viruses (Jackson *et al.*, 2016), and nucleocytoplasmic large DNA viruses (NCLDVs), as well as abundant bacteriophage (Hewson *et al.*, 2014). As such, due to the diversity of potential pathogens identified in affected organisms, and the inherent challenges of fulfilling Koch's postulates – even using molecular data (Fredericks and Relman, 1996) – in an invertebrate for which no cell cultures for viral isolation are available, it has become very difficult to point to any virus as a cause of SSW; hence additional mechanisms are being considered.

Sea star wasting

475

SSW has affected dozens of sea star species, spanning a broad phylogenetic breadth (Schiebelhut *et al.*, this issue). There are also numerous instances preceding the 2013-2014 outbreak and comparing the signs and etiology across such instances can be difficult (Hewson *et al.*, 2019). For this reason, it is not clear if the 2013-2014 outbreak is mechanistically related to prior or
480 ongoing observations of wasting; we do not know if the causes are the same across species or across events. A focus in this paper is on common structural traits of sea stars that may provide clues as to the signs and mortality that follow multiple potential causes, ranging from apparent pathogens to environmental stressors (Hewson *et al.*, 2018).

485

Sea stars, like other invertebrates, have a limited host response repertoire, so it would not be surprising that they respond to insults by manifesting lesions in the tissues that are most abundant (epidermis, MCT). As many of the affected species during the 2013-2014 SSW event are sympatric, they likely have similar exposure rates to the causal agent(s) of the disease (Eisenlord *et al.*, 2016), or environmental stressors (Dawson *et al.*, this issue). This indicates that
490 differences in the appearance or outcome among species may, instead, be driven by variations in the host immune response or other physical and physiological vulnerabilities. Understanding such variation may guide researchers in interpreting prior and ongoing asteroid wasting events that have similar signs or outcomes.

495

Regardless of why particular species were disproportionately affected by SSW over others, the die-off noted in 2013-2014 led to ecological shifts in echinoderm populations (Dawson *et al.*, this issue). For example, *Dermasterias imbricata* (the leather star) has increased in abundance at many sites (Montecino-Latorre *et al.*, 2016), and Kay *et al.* (2019) document “reciprocal abundance shifts” in the Salish Sea between the heavily affected *P. ochraceus*
500 and the less affected *Evasterias troschelii*. Other species, such as the sunflower star (*Pycnopodia helianthoides*), experienced sharp declines across its entire range, resulting in a nearly complete loss south of Oregon and functional depletion in much of its northern range

(Harvell *et al.*, 2019). *P. helianthoides* was recently listed by the International Union for Conservation of Nature as critically endangered, with greater than 90% of individuals lost (Gravem *et al.*, 2021). By contrast, it is unclear why species like *D. imbricata* persisted so successfully, but one hypothesis is that this species may have morphological or life history characteristics that affect disease severity (see Schiebelhut *et al.*, this issue). As an example of structural or ontogenetic variation that may be important, *Pisaster ochraceus* ochre morphs appear to be affected more than purple morphs (Raimondi *et al.*, 2007, Work *et al.*, 2021). This trait itself may be plastic (Harley *et al.*, 2006), serving as an indicator of other factors in the life of an asteroid that may be associated with severity of response.

Disease signs

While minor visible and pathological differences exist among host species, the onset of SSW is generally characterized by a progression of key visual signs. Sea stars first appear lethargic and uninterested in nearby prey (Kohl *et al.*, 2016), followed by an overall loss of body turgor and the ability to attach (Schultz *et al.*, 2016). Signs become more pronounced as arm rays twist and contort; and the sea stars quickly develop characteristic white lesions that mottle the epidermis (Figures 1 and 2; Hewson *et al.*, 2014, Eisenlord *et al.*, 2016, Bucci *et al.*, 2017).

Autotomy, or the detachment of arms (Figure 1) is a regular occurrence as lesions extend more deeply and wider into the tissues. Body wall tissues degrade and slough off, and the sea stars are often described as dissolving or “melting” from the outside inward as the animal dies (Eisenlord *et al.*, 2016). Progression of the disease may occur at a rapid pace; Menge *et al.* (2016) observed one star struggling to maintain its grip during a changing tide; 4 hours later they returned to find the animal in the final stages of disintegration and death.

Of course, SSW does not always progress linearly; there are cases documented where lesions form and then go away; limbs autotomize prior to lesions; or the lesions can form in the absence of deflation or arm twisting (Kohl *et al.*, 2016, Menge *et al.*, 2016). The progression can vary

between species or even between individuals within a species (Wares and Schiebelhut, 2016). The signs of SSW can also differ among species. For example, as opposed to the deflated *Pycnopodia* presented in Figure 1, *Dermasterias* can appear bloated and may have blue lesions rather than white. In some cases, the gonads are larger than in healthy animals and protrude out of the body through the lesions (SJW and Lesanna L. Lahner, Seattle Aquarium, WA, United States, oral communication, June 2020). This gonadal bloating is suggestive of a problem with osmotic homeostasis, and this hypothesis is also supported by the increase in chloride concentration detected in the coelomic fluid of affected sea stars (see Wahltinez et al., (2020) and section below on Immune responses in SSW).

The aforementioned outward signs of sea star wasting are not by themselves definitive proof that the animal has SSW. For example, loss of body turgor can occur normally in intertidal sea stars when exposed at low tide. Arm curling appears to be a normal response to a physical wound (Figure 3A,B), and may be observed in healthy, unwounded sea stars as well (Figure 3C). Arm autotomy also occurs following predation attempts (Ramsay *et al.*, 2000) and temperature stress (Pincebourde *et al.*, 2013). A related natural process of asexual reproduction, by splitting the body in half (fission) followed by regeneration, is seen in a few taxa such as *Coscinasterias muricata* (Byrne, 2020). Interestingly, SSW seems to be an extreme expression of these behaviors that occur typically in sea stars.

Histology

During the summer of 1991, an epizootic of SSW occurred in *Asterias forbesi*, with numerous animals washing up on the beaches along the coast of Long Island Sound and the Gulf of Maine (R.S., unpublished data). The animals had not autotomized arms, but did show flaccidity and extensive erosive, ulcerative, and penetrating lesions of the dorsal and dorsolateral surfaces of the arms. The ambulacral groove and tube feet were not affected, and no internal abnormalities were apparent. Gonads were shrunken in evaluated specimens; however, gonad maturity is seasonal and it is not clear if this is a sign associated with the SSW event. In terms of histology, internal organs showed no lesions while examination of

the dorsal epithelium showed areas of progressively, necrotizing dermatitis characterized by
565 an accumulation of coelomocytes in the superficial dermis and the base of the overlying
epithelium with vacuolation of epithelial cells and sloughing of the epithelial layer. Edema
and necrosis of the deeper dermal connective tissues and underlying ossicles occurred in the
ulcerated regions.

570 During the SSW epizootic that occurred on the Pacific Coast in 2013, histology was
evaluated for symptomatic and asymptomatic individuals of 13 asteroid species affected by
the event (A.L.N., unpublished data). Histologic lesions in *Pisaster ochraceus* and
Pycnopodia helianthoides were similar to those described above for *Asterias forbesi* (Figures
4 and 5; as in Bucci *et al.*, 2017). Some areas exhibited epidermal degeneration and necrosis
575 with ulceration. Areas of epidermal loss were associated with edema in the subjacent dermal
connective tissue and coelomocyte infiltrates. In addition to the epidermal and dermal
changes, coelomocyte aggregates were apparent within the coelomic cavity, vascular system,
and papulae with frequent adhesion of coelomocytes to the coelomic epithelial lining. There
were no primary lesions noted within the coelomic viscera or dermal ossicles and no
580 infectious agents were apparent (A.L.N., unpublished data). More recent studies looking at
experimental induction of SSW in *P. ochraceus* revealed that pathology involved a basal to
surface process starting with infiltration, breakdown of MCT, and lysis leading to epidermal
ulceration that had a greater effect on the purple than the ochre morphs. Indeed, ochre stars
survived longer, with more apparent and extensive signs of SSW, than those that are purple
585 (*Work et al.*, 2021).

The function and pathophysiological changes seen in the body wall in afflicted stars is
similar to that observed with the breakdown/autolysis of MCT during arm autotomy and at
the separation plane during asexual reproduction by fission in sea stars, processes that
590 are modulated by the nervous system (Mladenov and Burke, 1994, Wilkie, 2002; Byrne *et al.*,
2019). Hence, the changes in the body wall during SSW might, fundamentally, be a
neurological disease leading to inability to control softening of the MCT to the point that
total dissolution occurs. Additional research is needed to test this hypothesis and indeed to
better understand neural control of MCT tensility. Fuess *et al.* (2015) noted changes in

595 transcript abundance for genes associated with nervous system processes when comparing
wasting versus non-wasting stars. These differentially expressed transcripts included many
associated with MCT, supporting concern that the arm twisting and autotomy could be
related to dysregulation of this system.

600

Pedicellariae

As noted previously, a difficulty in defining disease in organisms like sea stars is to separate
abnormal condition from injury – this is not easy when body structures such as pedicellariae are
involved. In 2014, a mixed species group of sea stars including *Pisaster brevispinus*, *Pisaster*
605 *ochraceus*, *Evasterias troschelii*, *Dermasterias imbricata*, and *Orthasterias koehlerii* were placed
in tanks for a study on SSW. Detached pedicellariae from *Pisaster ochraceus* were noted
independently moving and penetrating the coelomic cavity of *D. imbricata*. This observation was
associated with clinical signs of SSW in *D. imbricata* (epidermal lesions, drooping limbs).

610 Removal of the pedicellariae reversed these clinical signs (SJW and Lesanna L. Lahner, Seattle
Aquarium, WA, United States, oral communication, June 2020). Though a sudden and dramatic
uptick in agonistic interactions would itself be of interest, these interactions became a focus of
study into mechanisms driving SSW.

615 To investigate the potential connection between pedicellaria attack and SSW, flow-through
holding tanks were set up, each with two adult *D. imbricata* and two adult *P. ochraceus*. Both
species initiated contact, and physical contact was required for pedicellariae transfer (i.e. defense
response) from *P. ochraceus* to *D. imbricata*, with the majority of the pedicellariae (88.9%)
transferred to the oral surface. In the area surrounding the transferred pedicellariae, the
620 ambulacral groove closed and tube foot dysfunction was noted. These signs immediately
resolved following manual removal of the pedicellariae (SJW and Lesanna L. Lahner, Seattle
Aquarium, WA, United States, oral communication, June 2020). These signs are consistent with
what has been noted after pedicellariae transfer from *Pisaster giganteus* and *Pisaster brevispinus*
to *Pycnopodia helianthoides* (Wobber, 1975) and from *Pisaster ochraceus* to *Solaster dawsoni*
625 (VanVeldhuizen and Oakes, 1981). The disuse of a limb following pedicellariae transfer could

be due to a toxin as seen in echinoid globiferous pedicellariae (Jensen, 1966, Chia, 1969), irritation, or another mechanism. Transfer of pedicellariae could represent a novel pathway for pathogen transmission or may predispose the recipient to infection through disruption of the epidermis or could contain an agent that causes the MCT to break down.

630

Immune responses in SSW

Very limited information is available on the immune responses of wasting stars. This is in part because of unclear associations of a putative pathogen across taxa – the densovirus initially proposed as a causal agent (Hewson *et al.* 2014) is now known to be as frequently found in asymptomatic individuals as in those showing signs of SSW. The overall class of viruses that have been primarily focused on (Sea Star associated Densovirus, SSaDV) is still potentially associated, in terms of viral load, with wasting disease in *Pycnopodia helianthoides* (Hewson *et al.* 2018) but does not correspond with the outcome of other challenged species – and the lack of mechanistic knowledge on the immune systems of the highly diverse group of species that are affected by wasting (Schiebelhut *et al.*, this issue). Coelomic fluid in *Pisaster ochraceus* with SSW had significantly increased chloride concentration, increased osmolality and coelomocyte count, suggestive of an osmotic breakdown (Wahlteinez *et al.*, 2020), when compared to clinically normal conspecifics (Wahlteinez *et al.*, 2019). The increased chloride concentrations and hypertonic coelomic fluid may be due, at least in part, to wound healing or inability to regulate osmolality of the coelomic fluid secondary to skin ulceration. An increase in chloride permeability was noted in sea star oocytes after wounding (Fein and Terasaki, 2005). In a more recent study, *P. ochraceus* manifesting experimental SSW showed a significant increase in a heretofore undescribed type of coelomocyte illustrating our very limited knowledge of sea star host responses to insults (Work *et al.*, 2021).

Another approach of great relevance to SSW is to measure the transcriptional profile in the coelomocytes of a wasting animal compared to healthy individuals. Fuess *et al.* (2015) used *Pycnopodia helianthoides* to explore the transcriptomic response of coelomocytes from sea stars injected with a <0.2 µm-tissue filtrate from an afflicted sea star. Within several days,

655

disease signs were seen in the test animals, but not in the control injected animals.

Remarkably, over 1000 immune-related genes were identified that were significantly changed in expression profile compared to the coelomocytes of the control animals. These
660 included pattern recognition receptors (PRRs) and other immune receptors, intracellular signaling, transcription factors, cytokines and growth factors, and immune effector genes, as well as genes involved in coagulation and the complement system, and homologues of genes that function in the vertebrate adaptive immune system.

665 The study of Fuess *et al.* (2015) raises a series of questions. Specifically, which of these genes are most important to SSW pathogenesis, and just how does this gene expression profile relate to host pathology at the cellular level? Also lacking in this experiment was a control for allorecognition reactions, such as a heat-killed tissue homogenate. Recall that the immune system of the sea star was defined early on by allorecognition and tissue rejection
670 (Smith *et al.*, 2010, Gordon, 2016a). Indeed, Aquino *et al.* (2021) argued that the $< 0.2 \mu\text{m}$ size fraction of sea star-derived filtered homogenate used in similar challenge studies may contain a broad array of dissolved organic matter, which ultimately may lead to allorecognition, but also may stimulate heterotrophic bacteria in subcuticular space or on/near surfaces, which ultimately may cause oxygen depletion and tissue degradation.

675

Regardless, the study of Fuess *et al.* (2015), as one of the very few contemporary studies on sea star molecular immunity, yielded valuable information on the transcriptomic response of sea stars to ‘non-self’ that could be used to develop more precise tests. An important follow up experiment could involve exposure of healthy stars to recombinantly expressed proteins of
680 candidate virus particles associated with SSW, as well as with specific antigens (PAMPs), environmental toxins, pedicellaria attack or isolated pathogens (e.g., bacteria) to test immune responses. The signature of SSW in these transcriptomic data has already been partially picked up in independent post-selection and heat stress experiments that are mapped to the genome of *Pisaster ochraceus* (Ruiz-Ramos *et al.*, 2020). The search for consilience across
685 several studies exploring wasting or stressors is a hopeful guide towards key mechanisms involved in the response.

As an alternative to whole animal studies, *in vitro* approaches have been performed by exposing purified immune cells (coelomocytes in absence of serum from the water vascular system) to various insults and then measuring in real time their responses using medium to high-throughput assays (Matranga *et al.*, 2002, Pinsino and Matranga, 2015). It may be more efficient to study responses in sea star larvae; immunocytes of sea star larvae share several similarities with adult coelomocytes, showing the capability for phagocytosis of cellular debris and foreign agents that invade blastocoel. Furukawa *et al.* (2009) showed that sea star larval immune cells phagocytose small foreign particles injected into the blastocoel within two hours. When small amounts are inserted, few cells respond. However, when larger particles are inserted into the blastocoel, multiple cells converge onto the site from within the blastocoel and undergo cell–cell fusion to form a syncytial multinucleated giant cell that encapsulates the aggregated particles. Similar syncytial formations have also been noted for adult phagocytes *in vitro* (Majeske *et al.*, 2013). The number of recruited immune cells is dependent on the amount and size of the foreign substance, indicating that the process of clearing the blastocoel is strictly regulated.

Candidate genes involved in this regulation appear to be two macrophage migration inhibitory factors (MIFs): ApMIF2 and ApMIF1 (Furukawa *et al.*, 2016). These immune effectors are evolutionarily ancient and highly conserved (Furukawa *et al.*, 2016). Therefore, the immune cell behaviors of chemotaxis, cytoskeletal modifications, and syncytia formation provide excellent nodes to investigate how the innate immune system functions generally, in a larva, and likely elements that are shared in an adult. Intriguingly, in sea cucumbers, the virus associated with the Acute Peristome Edema disease has been detected in both adults and 30 days old diseased larvae (Wang *et al.*, 2007). Currently, it is unknown whether embryos and larvae of sea stars are affected by SSW as well.

715

Potential agents associated with SSW

720 It is difficult to isolate the cause of SSW, which remains unknown in all spatial, temporal, and taxonomic context, from the proliferation of microbial or viral agents that may be found in compromised sea star tissues. There are often changes in microbial communities of affected individuals that indicate proliferation of several bacterial taxonomic groups and viruses (Hewson *et al.*, 2014, Hewson *et al.*, 2018, Lloyd and Pespeni, 2018, Hewson *et al.*, 2020a, Aquino *et al.*, 2021). Comparative viral metagenomic study of asymptomatic and wasting-
725 affected tissues revealed a proliferation of a virus (SSaDV or AaV-1; Hewson *et al.*, 2014). However, further study with redesigned qPCR primers targeting only SSaDV and excluding homologous non-SSaDV densovirus, revealed no association with wasting (Hewson *et al.*, 2018). More recent approaches also found no association between viruses and disease, and that densovirus form persistent infections in their hosts (Jackson *et al.*, 2020a, Jackson *et al.*,
730 2020b). More recent work tracking viral populations during progression of SSW in controlled laboratory experiments has revealed that wasting is associated with elevated richness of all viral genotypes, which may indicate a general proliferation of viruses in affected tissues or a down-regulation of the immune responses (Hewson *et al.*, 2020a).

735 Screening of SSW-affected tissues by histopathology in 2013 yielded no protist cells associated with lesions (Hewson *et al.*, 2014). Comparisons of fungal communities in *Odontaster validus* revealed no difference in associates between asymptomatic and SSW-affected states (Nunez-Pons *et al.*, 2018). While there have been no pan-eukarya surveys of protistan diversity in wasting asteroids, viral metagenomes also yield sequences of co-
740 extracted and amplified ribosome bound rRNAs (Hewson and Sewell, 2021). A survey of unicellular fungal and other eukaryotic 18S and 28S rRNAs in *Pisaster ochraceus* during wasting progression (Hewson *et al.*, 2020a) did not reveal any single OTU (operational taxonomic unit) consistently associated with wasting (Figure A1). In summary, there is little consistent evidence that infectious agents were responsible for the 2013-2014 mortality event
745 on the west coast of North America. Though the information available sometimes suggests association of viral load with signs of disease in certain populations (Hewson *et al.* 2018), the diverse and complex observations of SSW across the breadth of environmental and viral/prokaryotic communities that can be detected have made it challenging to narrow the likely sources.

750

Bacteria and Archaea also change in abundance and composition during SSW. It is believed that this is mainly due to stimulation of taxa that degrade organic matter released from decaying tissues. It is therefore unsurprising that the bacterial orders that associate with wasting tissues (Table A2) – notably Flavobacteriales (Lloyd and Pespeni, 2018),

755

Rhodobacteriales (Nunez-Pons *et al.*, 2018), and Campylobacteriales (Hoj *et al.*, 2018) are copiotrophs (Haggerty and Dinsdale, 2016). In terms of overall community structure, assemblages are different between healthy and diseased states (Nunez-Pons *et al.*, 2018). In *Acanthaster* sp., disease was associated with increasing dominance by fewer OTUs in the pyloric caeca but decreasing dominance (i.e. greater evenness) in the body wall tissues (Hoj

760

et al., 2018). Indeed, injection of bacterial culture media into *Acanthaster* sp. causes wasting-like lesions and animal death; isolated bacterial cultures from lesions included taxa in the genera *Vibrio* and *Photobacterium* (Rivera-Posada *et al.*, 2011a, Rivera-Posada *et al.*,

2011c). Moreover, healthy animals in contact with these infected animals also displayed signs of disease and died within 24 h, indicating that these first observations were not due to

765

a toxic shock after injection of the media. Because SSW is mainly defined by gross lesions that could have multiple etiologies, sorting out causes of sea star mortalities will require more systematic effort combining careful observations of tissues at the cellular level coupled with studies to relate molecular processes to tissue changes for each species.

770

Environment

Environmental effects on agents

775

Environmental conditions potentially affect the abundance and composition of microorganisms associated with SSW as both commensals and opportunists. Although much attention has been given to environmental correlates with SSW since 2013 (Eisenlord *et al.*, 2016, Kohl *et al.*, 2016, Menge *et al.*, 2016, Hewson *et al.*, 2018, Miner *et al.*, 2018, Aalto *et al.*, 2020), no link has been found between the microbiome composition in affected asteroids

780

and an environmental perturbation. Environmental stressors may be linked to dysbiosis, a

change in microbiome composition relative to the “normal” state (Egan and Gardiner, 2016); they could induce a proliferation of opportunistic pathogens when host tissues are compromised (Burge *et al.*, 2014). These stressors could lead to hypoxia through bacterial heterotrophic respiration (Haas *et al.*, 2011, Gregg *et al.*, 2013, Aquino *et al.*, 2021).

785 Environmental conditions could directly affect the ability of the host to respond to insults. Although viruses do not bear innate metabolism, their replication in tissues may be influenced by the intracellular conditions of their hosts. Thus, their proliferation may reflect breakdown in homeostatic properties as a consequence of environmental stressors in their poikilothermic hosts (Frakolaki *et al.*, 2018, Hewson *et al.*, 2020a). It is worth noting that
790 common taxa inhabiting asymptomatic asteroid specimens – including *Rhodobacter* (*Erythrobacter*), Flavobacteria, and Alteromonads – include some of the largest genome-bearing and fastest growth rate taxa in the picoplankton, which emphasizes their ability to rapidly adapt to changing conditions. The proliferation of these taxa during SSW may reflect their ability to rapidly assimilate organic matter from the environment surrounding asteroids
795 (Aquino *et al.*, 2021).

Environmental stressors

A common hypothesis is that anomalies in temperature may elicit or exacerbate SSW.

800 Although most studies have revealed that SSW is positively correlated to elevated sea surface temperatures (Bates *et al.*, 2009, Eisenlord *et al.*, 2016, Kohl *et al.*, 2016, Harvell *et al.*, 2019, Aalto *et al.*, 2020), other studies have found either no empirical relationship between the two variables (Hewson *et al.*, 2018), or concluded instead that a reduction in sea surface temperature (Menge *et al.*, 2016) is inducing or exacerbating the wasting disease. The
805 context-dependent relationship between temperature and SSW may result from an integration of both short- and long-term stressors, but may also instead indicate that the magnitude of the temperature anomaly induces wasting, regardless of the direction. In addition, some basic environmental variables can serve as proxies to environmental stressors, such as low-tide exposure and wave action (Dawson *et al.*, this issue). Early symptoms of SSW are similar to
810 those resulting from other sources of stress in sea stars, such as starvation, desiccation, or injury (Miner *et al.*, 2018, VanVolkom *et al.*, 2021). What is most intriguing about SSW is

that in many locations, predictive environments for sea star assemblages are changing rapidly (Kay *et al.*, 2019, Konar *et al.*, 2019).

815 Recently, Aquino *et al.* (2021) provided evidence that SSW could be related to microbial processes – driven by temperature and organic matter availability – at the animal-water interface. Microorganisms inhabiting the diffusive boundary layer around individual specimens may respond to inputs from surrounding waters. In marine environments, a large proportion of this organic matter is in the form of phytoplankton exudates (Ogawa and
820 Tanoue, 2003). Aquino *et al.* (2021) observed correspondence between chlorophyll *a* peak (and decline) and SSW mass mortality at a field site, and Hewson (2021) noted correspondence between SSW and upwelling at several sites, so it is plausible that SSW is influenced by water column processes such as stratification, nutrient availability, and temperature. Linking SSW to biological oceanographic phenomena may also help explain the
825 multi-host nature of SSW and the apparent discontinuous geographic appearance of SSW in 2013-2014, and seasonal nature of the condition in years since (Bates *et al.*, 2009). We note here that mass mortalities of echinoderms from other possible environmental mechanisms, such as the toxins from harmful algal blooms, can be contrasted in parts of California (Jurgens *et al.*, 2015) with those of SSW. Related toxins were evaluated in sea stars, with no
830 relationship to wasting status (Hewson *et al.*, 2018).

Finally, asteroids produce chemical compounds under stress that may induce wasting-like conditions in grossly normal specimens when challenged via direct injection. In response to short exposure (45-90 sec) to heat stress, asteroids release proteinaceous compounds within
835 their coelomic fluid that cause almost immediate body wall softening, limb autotomy, and death when injected into healthy specimens (Chaet, 1962, Mladenov *et al.*, 1989). Although these factors were obtained by immersing specimens at 76°C, an extreme that these species would never experience, these results illustrate that stressed asteroids have the potential to affect the health of surrounding individuals. Because these compounds also occur within the
840 material tested during challenge experiments (Hewson *et al.*, 2014, Bucci *et al.*, 2017, Hewson *et al.*, 2018, Aquino *et al.*, 2021), it is possible that such factors may have been

responsible for the observation of apparent transmission, and bring us full circle with questions about changing density and abundance.

845

Contrasts with other marine diseases and mass mortality events

Unfortunately, SSW is not a unique phenomenon in terms of huge, sudden impacts on marine
850 communities. Our growing recognition of mass mortality events in the ocean (e.g., coral
bleaching and death (Hughes *et al.*, 2018a, Hughes *et al.*, 2018b)) dovetails with the
associated recognition of how climate change will ultimately affect many taxa, e.g, dolphins
(Duignan *et al.*, 2020), manatees (Hardy *et al.*, 2019), and other invertebrates (Petraitis and
Dudgeon, 2020). The difficulties in pinpointing a cause for disease are increased when the
855 environmental background is rapidly changing (Burge *et al.*, 2014, Groner *et al.*, 2016, Tracy
et al., 2019).

Extraordinary waves of disease are now more frequently the focus of marine research
(Harvell *et al.*, 1999). Multiple outbreaks of seagrass wasting are being explored in the
860 western Atlantic, northeastern Pacific, and elsewhere (Short *et al.*, 1987, Muehlstein *et al.*,
1991, Sullivan *et al.*, 2018). A series of devastating diseases have reduced coral abundance in
the Caribbean – the most recent wave being ‘stony coral tissue loss’ disease (Aeby *et al.*,
2019, Landsberg *et al.*, 2020) – concomitant with increasing frequency of climate-driven
bleaching. Sponges (Olson *et al.*, 2006), shrimp (Frischer *et al.*, 2017), and salmon (Asche *et*
865 *al.*, 2009) were also affected by major disease outbreaks, that together with climate change,
are shifting entire ecosystems (Tracy *et al.*, 2019).

Focusing on echinoderms, we are left to look back on previous mass mortality events for lessons
of what comes next. Echinoids are known to be affected by “bald sea urchin disease” (Johnson,
870 1971). They present symptoms such as body wall lesions where spines, tube feet, pedicellaria
and epidermis are lost. Sometimes necrosis leads to the perforation of the test and the death of
the animals. The bacteria later detected in this disease were found to be opportunistic instead of

specific (Becker *et al.*, 2008). In the case of *Diadema antillarum*, no mechanism is known for the mass mortality in the Caribbean; current study is focused on how and where it is recovering its numbers (Myhre and Acevedo-Gutiérrez, 2007, Lessios, 2016). In the Sea of Cortez, the recovery of *Heliaster kubiniji* has been spatially variable (Eckert *et al.*, 2000).

Currently, some species have not recovered from SSW (Gravem *et al.*, 2021) while others are returning to their former abundance (Dawson *et al.*, this issue) but with localized die-offs from this unknown cause (Moritsch and Raimondi, 2018). We know there have been similar outbreaks and apparent recovery (Hewson *et al.*, 2018), and we know that asteroids cycle demographically over long-time scales (Uthicke *et al.*, 2009). Finally, there are current studies documenting new losses in holothuroids (Hewson *et al.*, 2020b), and the ecological effects of loss are likely to have substantial effects on coastal benthic communities (Harrold and Pearse, 1987, McPherson *et al.*, 2021).

Conclusions and future directions

Sea star wasting, characterized by behavioral changes, body lesions, and arm autotomy, has triggered periodic mass mortality events across a broad range of sea star species – and possibly with similar mechanisms across disparate events. To better characterize the etiology, the epidemiology, and the effect of SSW, it is essential to build a community of investigators that can share knowledge, samples, and protocols (Table A3, and Supplement S1, available online) and monitor the corresponding environmental parameters around the world.

Understanding SSW – especially the most recent iterations – is challenging because it is now affecting so much diversity, the ecological consequences of which have yet to be fully appreciated. Our primary challenge is that the basic biology of the sea stars is surprisingly poorly known, making it difficult to directly compare healthy and symptomatic animals. This challenge is further compounded by studying these dynamics in a rapidly changing environment and with increased scrutiny of species and populations that have previously not been closely monitored. A deeper knowledge of the basic biology, health, disease, and life stage dynamics and of the growth of sea stars with insights from a range of disciplines will

905 be essential to understand how and why these animals are so profoundly affected by or react
to SSW conditions.

Acknowledgements

910 The virtual workshop supporting development of this project and participation of JPW,
MND, LMS, PD, IH was funded by the National Science Foundation (NSF) grants OCE-
1737091, OCE-1737381, OCE-1737127. We thank the reviewers for their comments and
915 suggestions. We appreciate the encouragement of our many colleagues who have shared their
previously unpublished data or results, and photographers who have allowed us to use their
images. Any use of trade, firm, or product names is for descriptive purposes only and does
not imply endorsement by the U.S. Government.

920 **Figure legends:**

Figure 1: Examples of sea star species affected by SSW. The left column represents animals with no or with early symptoms. The right column represents animals with late symptoms.

925 (A) *Pycnopodia helianthoides*, the left image shows terminal stages of SSW, Photo credit:

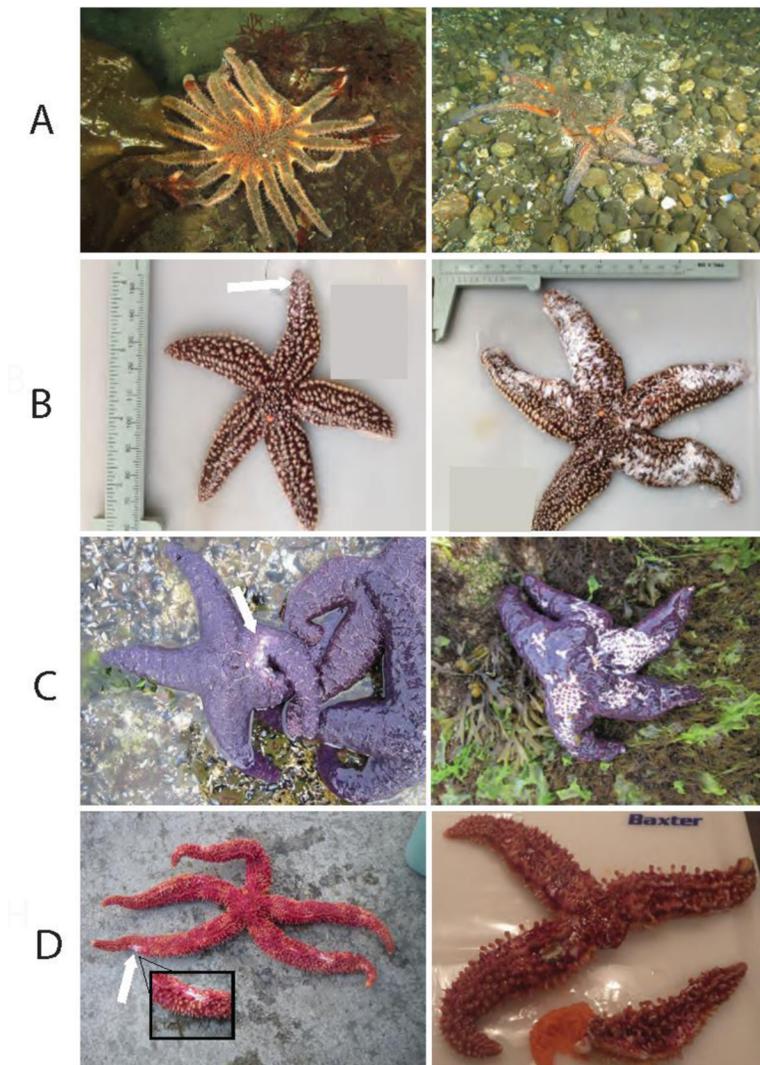
Mark Nayer (B) *Asterias forbesi*, the arrow on the left image shows a small lesion. (C)

Pisaster ochraceus, the arrow on the left image shows a small lesion, Photo credit: Melissa

Miner (D) *Orthasterias*, the arrow on the left image shows a small lesion, Photo credit: Feiro

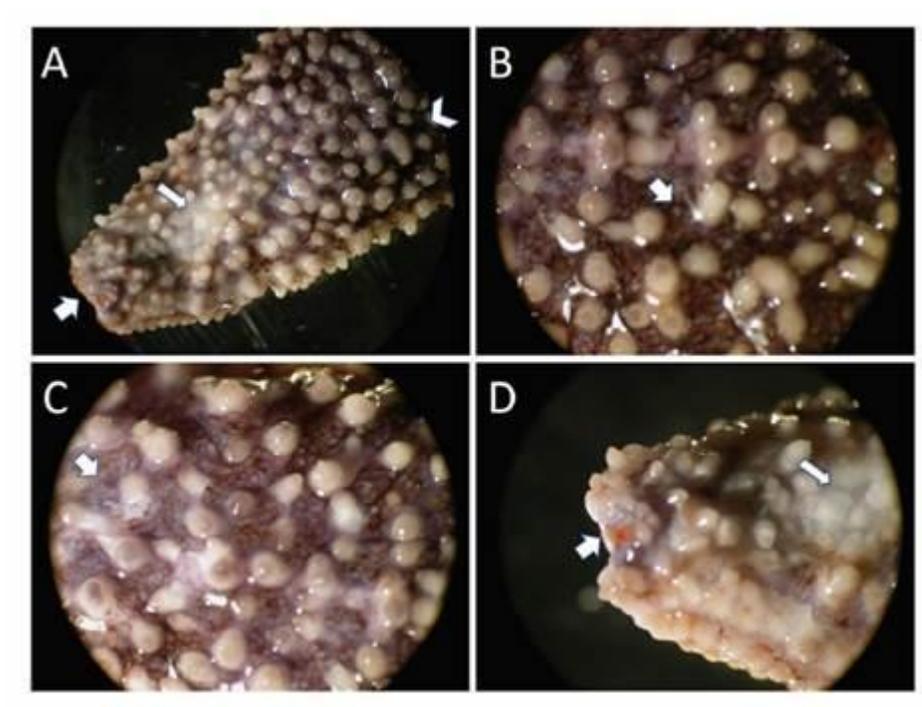
Marine Life Center (left).

930



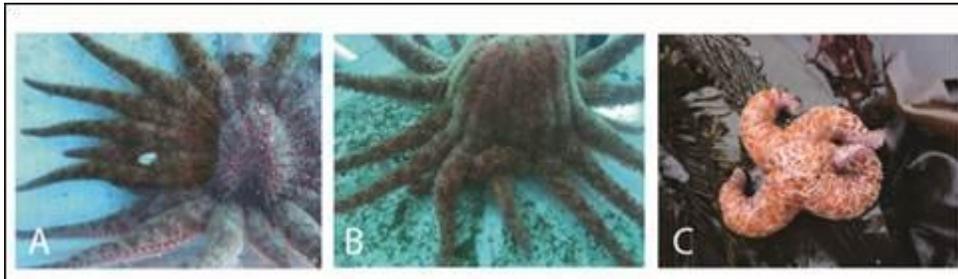
935 **Figure 2:** Progression of gross lesions in the aboral side of sea star wasting in *Asterias*
forbesi. (A) Arm tip of an affected sea star showing a progression from healthy-looking
tissue (chevron, top right) to broad areas of discoloration and loss of cuticle and epithelial
tissue (narrow arrow) and severe ulcers leading to loss of tissue at the tip of the arm and
exposure of underlying ossicle and the tip of the radial canal (broad arrow). (B) Close up of
940 early stage lesion with small pinpoint white lesions (broad arrow) and early degradation of
the spines. (C) Close up of a more advanced lesion showing further development of pinpoint
white lesions (broad arrow) and spine degradation. (D) Close up of the tip of the arm in (A)
showing advanced lesions with total loss of epithelium (narrow arrow) and exposure of the
underlying tissues and ossicles and the tip of the radial canal (broad arrow).

945



950

Figure 3: Arm twisting is a behavior that can be present in both healthy and wasting sea stars, and thus cannot always be attributed to SSW. (A,B) *Pycnopodia helianthoides*. (A) shows the wound, and (B) shows how the non-wasting sea star covers this wound by twisting its arms. (C) *Pisaster ochraceus*, The twisting behavior is here observed in a sea star that showed no other signs of SSW. Photo credits: A,B–Julia Kobelt; C–Mike Dawson.



960

965

970 **Figure 4:** Early lesions of the dorsal epithelium of an *Asterias forbesi* with SSW. (A)
vacuolation and necrosis of epithelial cells with influx of hemocytes (arrow heads) and
disruption of the cuticle; (B) edema of underlying connective tissues; (C) necrosis of muscle
and associated underlying connective tissues. (Bar = 32 μ m).

975

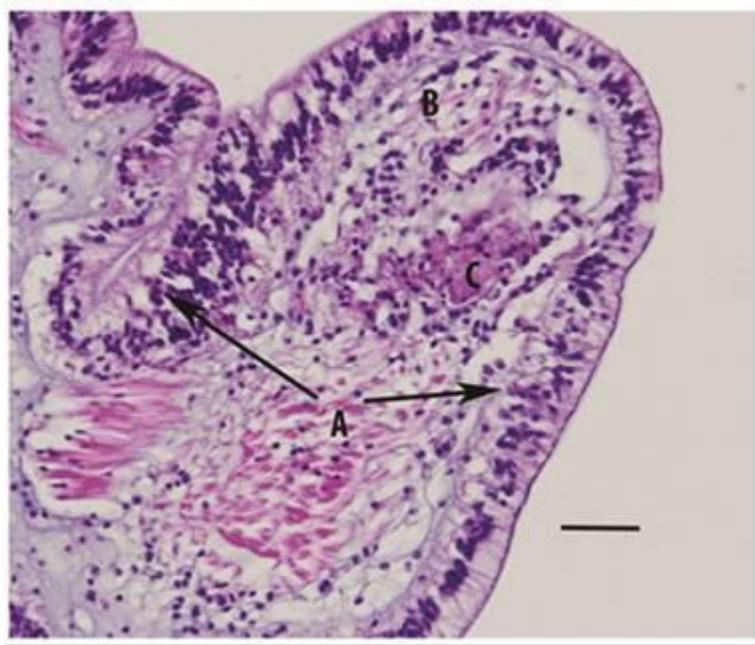
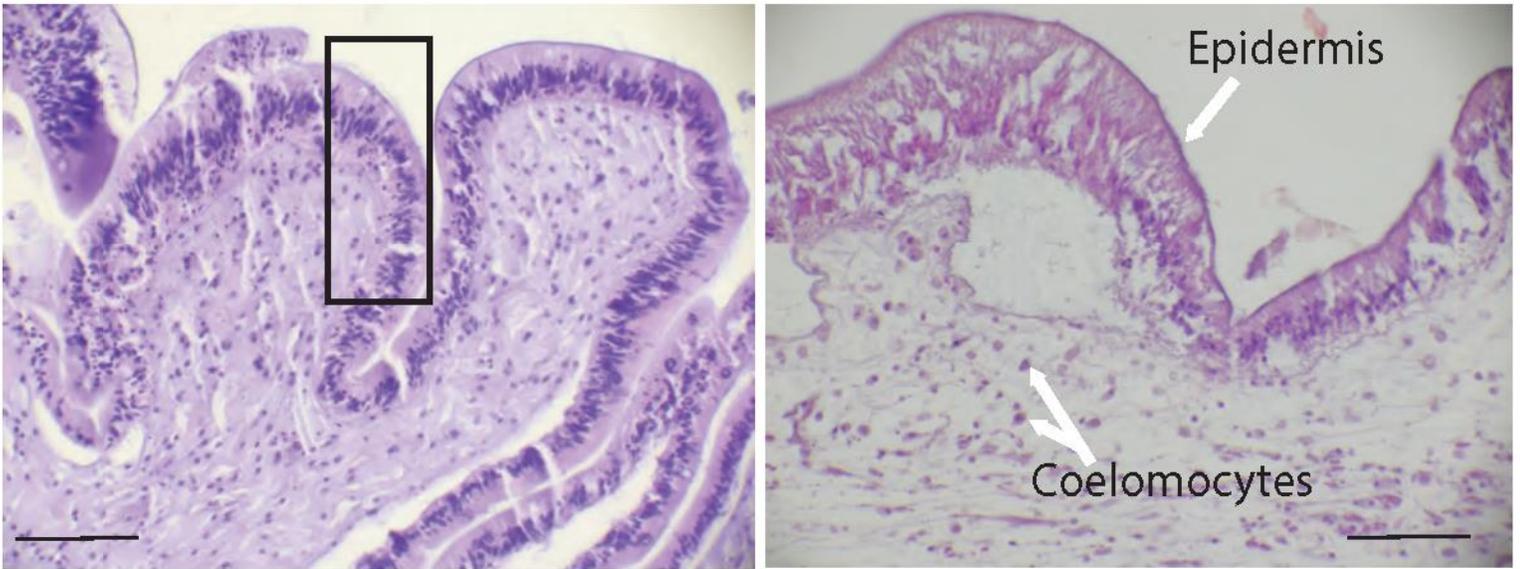


Figure 5: Common lesions noted in the epidermis and dermis. On the left (*Pisaster ochraceus*), within the box, there is multifocal epidermal degeneration and lytic necrosis characterized by a loss of cell distinction, nuclear condensation and fragmentation (karyorrhexis). On the right (*Pycnopodia helianthoides*), there is coagulation type epidermal necrosis characterized by loss of cellular detail, particularly nuclear and cell margin distinction with increased uptake of eosin stain (pink staining) by the cytoplasm. Both types of lesions were associated with microscopic epidermal loss or ulceration in some specimens. The right image also shows the two most common lesions within the dermis: separation of the connective tissue fibers by edema and infiltrates of coelomocytes consistent with inflammation. A low instance of dermal degeneration and necrosis also occurred. Scale bars are 50µm.



1005 **Appendix Table 1:** Cultivated bacterial isolates from grossly normal asteroids. [1] (Narita *et al.*, 1987), [2] (Rivera-Posada *et al.*, 2011b), [3] (Hewson *et al.*, 2018), [4] (Luo *et al.*, 2013), [5] (Beleneva and Zhukova, 2009), [6] (Choi *et al.*, 2010a), [7] (Choi *et al.*, 2003), [8] (Choi *et al.*, 2010b), [9] (Ivanova *et al.*, 2004), [10] (Zhang *et al.*, 2016).

1010

Phylum	Class	Order	Family	Genus	Reference
Proteobacteria	<u>Gammaproteobacteria</u>	<u>Vibrionales</u>	<u>Vibrionaceae</u>	<u>Vibrio</u> , <u>Photobacterium</u> , <u>Alivibrio</u>	[1,2,3]
		<u>Alteromonadales</u>	<u>Pseudoalteromonadaceae</u>	<u>Pseudoalteromonas</u>	[3,4]
			<u>Shewanellaceae</u>	<u>Shewanella</u>	[3,5]
			<u>Cobwelliaceae</u>	<u>Cobwellia</u>	[6]
			<u>Alteromonadaceae</u>	<u>Marinobacter</u>	[7]
		<u>Pseudomonadales</u>	<u>Pseudomonadaceae</u>	<u>Pseudomonas</u>	
			<u>Moraxellaceae</u>	<u>Psychobacter</u>	[7]
		<u>Oceanospirillales</u>	<u>Halomonadaceae</u>	<u>Halomonas</u>	[5]
			<u>Hahellaceae</u>	<u>Kistomonas</u>	[8]
	<u>Alphaproteobacteria</u>	<u>Rhodobacterales</u>	<u>Rhodobacteriaceae</u>	<u>Sulfitobacter</u> , <u>Lutimaribacter</u> , <u>Paracoccus</u>	[7,9,10]
		<u>Sphingomonadales</u>	<u>Sphingomonadaceae</u>	<u>Erythrobacter</u>	[7,9]
	<u>Betaproteobacteria</u>	<u>Rhodocyclales</u>	<u>Rhodocyclaceae</u>	<u>Zoogaea</u>	[7]
Actinobacteria	Actinobacteria	<u>Corynebacteriales</u>			[5]
		<u>Actinomycetales</u>	<u>Microrcoccaceae</u>	<u>Arthrobacter</u> , <u>Microbacterium</u> , <u>Kocuria</u>	[5,7]
Firmicutes	Bacilli	<u>Bacillales</u>	<u>Bacillaceae</u>	<u>Bacillus</u>	[5,7]
			<u>Staphylococcaceae</u>	<u>Staphylococcus</u>	[5,7]
Bacteroidetes	Flavobacteria	<u>Flavobacteriales</u>	<u>Flavobacteraceae</u>	<u>Psychoserpens</u>	[3,5]
Planctomycetes					[5]

Appendix Table 2: Dominant bacterial taxa represented in cultivation-independent surveys.

Data derived from [1] (Nakagawa *et al.*, 2017), [2] (Hoj *et al.*, 2018), [3] (Jackson *et al.*, 2018), [4] (Lloyd and Pespeni, 2018), [5] (Nunez-Pons *et al.*, 2018) and [6] (Hewson *et al.*, 2018). *** indicates taxon comprised >50% of sequences surveyed; ** indicates taxon comprised > 20% of sequences surveyed; * indicates taxon comprised < 20 % of sequences surveyed.

1015

1020

Phylum	Class	Family	<i>Asterias amurensis</i> [1]	<i>Acanthaster planci</i> [2]	<i>Pisaster ochraceus</i> [3]	<i>Odontaster balidus</i> [4]	Multiple species [5]	
Proteobacteria	Alphaproteobacteria	Rhodobacteraceae	*	**	*	**	***	
		Caulobacteriales		*			*	
		Rhizobacteriales		*	*		**	
		Sphingomonadales		*			*	
		Rhodospirillales					*	
			Rickettsiales				*	
	Betaproteobacteria		Nitrosomonadaceae	*				
			Burkholderiales		*		*	*
			Neisseriales		*			
	Gammaproteobacteria		Thiotrichales	***				
			Vibrionaceae	*	*	*		
			Alcanivoracaceae	*				
			Alteromonadales		*	**	*	**
			Enterobacteriales		*			**
			Oceanospirillales		***	*		*
			Pseudomonadales		*	*	*	*
			Xanthomonadales		*			*
			Chromatiales					*
			Deltaproteobacteria	SAR324				
	Epsilonproteobacteria		Helicobacteraceae	***				
		Campylobacteraceae	*	*	*			
	Oligoflexia						*	
Bacteroides	Flavobacteriia	Flavobacteriaceae	**	**	**	**		
		Cryomorphaceae	*					
	Bacteroides			*				
	Sphinzobacteriia			*				
		Cytophazales				*		
Cyanobacteria	Cyanophaceae	Oscillatoriaceae		*				
Firmicutes	Bacilli	Lactobacillales		*				
	Clostridia	Clostridiales		*		*		
	Bacillia	Bacillales				*	*	
Spirochaetes	Spirochetia			***	***		**	
Tenericutes	Mollicutes		***	***			***	
Verrucomicrobia	Verrucomicrobiae	Verrucomicrobiaceae	*					
Actinobacteria	Actinobacteria	Actinomycetaceae		*			*	
		Corynebacteriales				***		

Appendix Table 3: How to store the samples? Appropriate preservation and storage of the samples are key elements in these studies, and consistent sampling and storage are critical for current and future studies of wasting. In particular, we would like to offer simple sampling and storage protocols so that researchers anywhere can take advantage of wasting (and non-wasting) specimens to be most useful to the research community at large. In the online supplement, we provide best practices for characterizing and analyzing individuals that are encountered with wasting-like symptoms. *Indicated significant loss of target organism or intact particles suitable for metagenomic approaches; ** the effects of TriZol with freezing on the stability of microbial nucleic acids has not been assessed. In the cost column, the expected price of the method is described from the cheapest (\$) to the most expensive (\$\$\$). The abbreviation IATA means International Air Transport Association.

Preservation Method	Cost	IATA Shipping Regulation	Compatibility for Host Studies				Compatibility for Molecular Microbial Studies			
			RNA	DNA	Microscopy / TEM	Histology	RNA Viruses	DNA Viruses	Bacteria and Archaea	Eukaryotic Microorganisms
Freezing at -20°C	\$	Dry Ice; Packing Instruction 904	N	Y	N	N	N	Y	Y	Y
Freezing at -80°C	\$\$\$	Liquid N2 not permitted; Dry shippers exemption A152	Y	Y	N	N	Y	Y	Y	Y
RNA Later (= Freezing)	\$\$	None	Y	N	N	N	N*	N	Y	Y
Trizol (= Freezing)	\$\$	Corrosive liquid, n.o.s., hazard class 8	Y	Y	N	N	**	**	**	**
Formalin	\$	DG Class II	N	N	Y	Y	N	N	N	N
≥95% Ethanol	\$	Exempted Quantities	Y*	Y	N	N	Y*	Y	Y	Y

References

1040

Aalto, E. A., K. D. Lafferty, S. H. Sokolow, R. E. Grewelle, T. Ben-Horin, C. A. Boch, P. T.

Raimondi, S. J. Bograd, E. L. Hazen, M. G. Jacox et al. 2020. Models with environmental drivers offer a plausible mechanism for the rapid spread of infectious disease outbreaks in marine organisms. *Sci. Rep.* **10**:5975.

1045

Aeby, G. S., B. Ushijima, J. E. Campbell, S. Jones, G. Williams, J. L. Meyer, et al. 2019.

Pathogenesis of a tissue loss disease affecting multiple species of corals along the Florida reef tract. *Front. Mar. Sci.* **6**:18.

Alupay, J. S. 2013. Characterization of Arm Autotomy in the Octopus, *Abdopus aculeatus* (d'Orbigny, 1834). C Berkeley, ProQuest ID: Alupay_berkeley_0028E_14043. Merritt ID: ark:/13030/m56h7mt8.

1050

Anderson, J. M. 1953. Structure and function in the pyloric caeca of *Asterias forbesi*. *Biol. Bull.* **105**:47-61.

Anderson, J. M. 1954. Studies on the cardiac stomach of the starfish *Asterias forbesi*. *Biol. Bull.* **107**:157-173.

Anderson, J. M. 1959. Studies on the cardiac stomach of a starfish *Patira miniata*. *Biol. Bull.* **117** 185-201.

1055

Aquino, C. A., R. M. Besemer, C. M. DeRito, J. Kocian, I. R. Porter, P. T. Raimondi, J. E. Rede, L. M. Schiebelhut, J. P. Sparks, J. P. Wares et al. 2021. Evidence that microorganisms at the animal-water interface drive sea star wasting disease. *Front. Microbiol.* **11**:19.

Asche, F., H. Hansen, R. Tveterås, and S. Tveterås. 2009. The salmon disease crisis in Chile. *Mar. Resour. Econ.* **24**:405-411.

1060

Bang, F. B. 1982. Disease processes in sea stars: A Metchnikovian challenge. *Biological Bulletin* **162**:135-148.

- Barbaglio, A., S. , C. Tricarico, D. Di Benedetto, A. P. Fassini, A. R. Lima, C. C. Ribeiro, M. Ribeiro, F. Sugni, L. Bonasoro, and E. A. Wilkie. 2013.** The smart connective tissue of echinoderms: A materializing promise for biotech applications. *Cahiers de Biologie Marine*:713-720.
- Barbaglio, A., S. Tricarico, A. Ribeiro, C. Ribeiro, M. Sugni, C. Di Benedetto, I. Wilkie, M. Barbosa, F. Bonasoro, and M. D. Candia Carnevali. 2012.** The mechanically adaptive connective tissue of echinoderms: its potential for bio-innovation in applied technology and ecology. *Mar. Environ. Res.* **76**:108-113.
- Bates, A. E., B. J. Hilton, and C. D. Harley. 2009.** Effects of temperature, season and locality on wasting disease in the keystone predatory sea star *Pisaster ochraceus*. *Dis. Aquat. Organ.* **86**:245-251.
- Becker, P. T., E. Egea, and I. Eeckhaut. 2008.** Characterization of the bacterial communities associated with the bald sea urchin disease of the echinoid *Paracentrotus lividus*. *Journal of Invertebrate Pathology* **98**:136-147.
- Beleneva, I. A., and N. V. Zhukova. 2009.** Seasonal dynamics of cell numbers and biodiversity of marine heterotrophic bacteria inhabiting invertebrates and water ecosystems of the Peter the Great Bay, Sea of Japan. *Microbiology* **78**:369-375.
- Binyon, J. 1961.** Salinity tolerance and permeability to water of the starfish *Asterias rubens*. *J. Mar. Biol. Ass. UK* **41**:161-174.
- Binyon, J. 1962.** Ionic regulation and mode of adjustment to reduced salinity of the starfish *Asterias rubens*. *L. J. Mar. Biol. Assoc. United Kingdom* **42**: 49.
- Binyon, J. 1972.** *Physiology of Echinoderms*, Oxford.
- Blake, D. B. 1989.** *Asteroidea: Functional morphology, classification and phylogeny*. A.A. Balkema, Rotterdam.
- Bucci, C., M. Francoeur, J. McGreal, R. Smolowitz, V. Zazueta-Novoa, G. M. Wessel, and M. Gomez-Chiarri. 2017.** Sea Star Wasting Disease in *Asterias forbesi* along the Atlantic Coast of North America. *PLoS One* **12**:e0188523.

- Burge, C. A., C. Mark Eakin, C. S. Friedman, B. Froelich, P. K. Hershberger, E. E. Hofmann, L. E. Petes, K. C. Prager, E. Weil, B. L. Willis et al. 2014.** Climate change influences on marine infectious diseases: implications for management and society. *Ann. Rev. Mar. Sci.* **6**:249-277.
- 1090
- Burt, J. M., M. T. Tinker, D. K. Okamoto, K. W. Demes, K. Holmes, and A. K. Salomon. 2018.** Sudden collapse of a mesopredator reveals its complementary role in mediating rocky reef regime shifts. *Proc. Biol. Sci.* **285**:20180553.
- Byrne, M. 1995.** Change in larval morphology in the evolution of benthic development by *Patiriella exigua* (Asteroidea: Asterinidae), a comparison with the larvae of *Patiriella* species with planktonic development. *Biol. Bull.* **188**:293-305.
- 1095
- Byrne, M. 1999.** *Echinodermata*. Academic Press, New York.
- Byrne, M. 2013.** *Asteroid evolutionary developmental biology and ecology*, Hopkins Press, Baltimore.
- Byrne, M. 2020.** The link between autotomy and CNS regeneration: Echinoderms as non-model species for regenerative biology. *Bioessays* **42**:e1900219.
- 1100
- Byrne, M., A. Cerra, i. T. Nishigak, and M. Hoshi. 1997.** Infestation of the testes of the Japanese sea star *Asterias amurensis* by the ciliate *Orchitophrya stellarum*: A caution against the use of this ciliate for biological control. *Dis. Aquat. Org.* **28**:235-239.
- Byrne, M., T. Cerra, T. Nishigaki, and M. Hoshi. 1998.** *Male infertility: A new phenomenon affecting Japanese populations of the sea star Asterias amurensis (Asteroidea) due to the introduction of the parasitic ciliate Orchitophryastellarum to Japan*, In: Mooi R, Telford M (eds) *Echinoderms*: San Francisco. Balkema, Rotterdam.
- 1105
- Byrne, M., F. E. W. Rowe, L. M. Marsh, C. L. Mah, and T.D. 2017.** *Class Asteroidea. In: Australian Echinoderms: Biology Ecology and Evolution.*
- 1110
- Byrne M, Mazzone F, Elphick MR, Thorndyke MC, and Cisternas P. 2019.** Expression of the neuropeptide SALMFamide-1 during regeneration of the seastar radial nerve cord following arm autotomy. *Proc. Roy. Soc. B*:20182701

- Cameron, C. B., and C. D. Bishop. 2012.** Biomineral ultrastructure, elemental constitution and genomic analysis of biomineralization-related proteins in hemichordates. *Proc R Soc B.* **279**:3041–3048.
- 1115 **Campbell, A. C., and P. S. Rainbow. 1977.** The role of pedicellariae in preventing barnacle settlement on the sea-urchin test. *Mar. Behav. Physiol.* **4**:253-260.
- Cepede, C. 1907.** La castration parasitaire des etoiles de mer males par un nouvel infuoire astome: *Orchitophrya stellarum*, n.g., n. sp. *CR Hebd Seances Academie Sciences Paris* **145**:1305-1306.
- Chaet, A. B. 1962.** A toxin in the coelomic fluid of scalded starfish (*Asterias forbesi*). *Exper. Biol. Med.* 1120 **109**:791-794.
- Chia, F., and H. Amerongen. 1975.** On the prey-catching pedicellariae of a starfish, *Stylasterias forreri* (de Loriol). *Can. J. Zool.* **53**:748-755.
- Chia, F., and R. Koss. 1994.** *Asteroidea*, Wiley-Liss, New York.
- Chia, F. S. 1968.** Embryology of a brooding starfish *Leptasterias hexactis* (Stimpson). *Acta Zoologica* 1125 **49**:321-354.
- Chia, F. S. 1969.** Response of the globiferous pedicellariae to inorganic salts in three regular echinoids. *Ophelia* **6**:203-210.
- Chia, F. S., C. Oguro, and M. Komatsu. 1993.** Sea-star (asteroid) development. *Oceanography and Marine Biology: An Annual Review* **31**:223-257.
- 1130 **Choi, E. J., H. C. Kwon, H. Y. Koh, Y. S. Kim, and H. O. Yang. 2010a.** *Colwellia asteriadis* sp. nov., a marine bacterium isolated from the starfish *Asterias amurensis*. *Int. J. Syst. Evol. Microbiol.* **60**:1952-1957.
- Choi, E. J., H. C. Kwon, Y. C. Sohn, and H. O. Yang. 2010b.** *Kistimonas asteriae* gen. nov., sp. nov., a gammaproteobacterium isolated from *Asterias amurensis*. *Int. J. Syst. Evol. Microbiol.* **60**:938-943.
- 1135 **Choi, G.-G., O.-H. Lee, and G.-H. Lee. 2003.** The diversity of heterotrophic bacteria isolated from intestine of starfish (*Asterias amurensis*) by analysis of 16S rDNA Sequence *The Korean Journal of Ecology* **26**:307-312.

- 1140 **Cintra-Buenrostro, C. E., H. Reyes-Bonilla, and M. D. Herrero-Perezrul. 2005.** Oceanographic conditions and diversity of sea stars (Echinodermata: Asteroidea) in the Gulf of California, Mexico. *Rev Biol Trop* **53 Suppl 3**:245-261.
- Claereboudt, M., and C. Bouland. 1994.** The effect of parasitic castration by a ciliate on a population of *Asterias vulgaris*. *Journal of Invertebrate Pathology* **63**:172-177.
- Cobb, J. 1987.** *Neurobiology of the echinodermata*. New York: Plenum Press.
- 1145 **Cobb, J. L. 1978.** An ultrastructural study of the dermal papulae of the starfish, *Asterias rubens*, with special reference to innervation of the muscles. *Cell Tissue Res* **187**:515-523.
- Cramer, K. L., J. B. C. Jackson, M. K. Donovan, B. J. Greenstein, C. A. Korpanty, G. M. Cook, and J. M. Pandolfi. 2020.** Widespread loss of Caribbean acroporid corals was underway before coral bleaching and disease outbreaks. *Sci Adv* **6**:eaax9395.
- 1150 **Dearborn, J. H., K. C. Edwards and D. B. Fratt. 1991.** Diet, feeding behavior, and surface morphology of the multi-armed Antarctic sea star *Labidiaster annulatus* (Echinodermata: Asteroidea). *Mar. Ecol. Prog. Ser.* **77**:65-84.
- Diehl, W. J., and J. M. Lawrence. 1984.** The effect of salinity on coelomic fluid osmolyte concentration and intracellular water content in *Luidia clathrata* (Say) (Echinodermata: Asteroidea). *Comp. Biochem. Physiol. Part A Physiol.* **79**:119-126.
- 1155 **Duggins, D. O. 1983.** Starfish predation and the creation of mosaic patterns in a kelp-dominated community. *Ecology* **64**:1610-1619.
- Duignan, P. J., N. S. Stephens, and K. Robb. 2020.** Fresh water skin disease in dolphins: a case definition based on pathology and environmental factors in Australia. *Sci Rep* **10**:21979.
- 1160 **Dungan, M. L., T. E. Miller, and D. A. Thomson. 1982.** Catastrophic decline of a top carnivore in the Gulf of California rocky intertidal zone. *Science* **216**:989-991.
- Eckert, G. L., J. M. Engle, and D. J. Kushner. 2000.** Sea star disease and population declines at the Channel Islands. *Proc. 5th Calif. Islands Symp*:390-393.

- Egan, S., and M. Gardiner. 2016.** Microbial dysbiosis: Rethinking disease in marine ecosystems. *Front. Microbiol.* 7:991.
- 1165 **Eisenlord, M. E., M. L. Groner, R. M. Yoshioka, J. Elliott, J. Maynard, S. Fradkin, M. Turner, K. Pyne, N. Rivlin, R. van Hooidek et al. 2016.** Ochre star mortality during the 2014 wasting disease epizootic: role of population size structure and temperature. *Phil. Trans. R. Soc. Lond. B Biol. Sci.* 371:20150212.
- 1170 **Ellington, W. R., and J. M. Lawrence. 1974.** Coelomic fluid volume regulation and isosmotic intracellular regulation by *Luidia clathrata* (Echinodermata: Asteroidea) in response to hyposmotic stress. *Biol Bull* 146:20-31.
- Emlet, R. B., L. R. McEdward, and R. R. Strathmann. 1987.** Echinoderm larval ecology viewed from the egg. *Echinoderm Stud* 2:55-136.
- 1175 **Emson, R. H., and C. M. Young. 1994.** Feeding mechanism of the brisingid starfish *Novodinia antillensis*. *Mar. Biol.* 118:433-442.
- Erlinger, R., U. Welsch, and J. E. Scott. 1993.** Ultrastructural and biochemical observations on proteoglycans and collagen in the mutable connective tissue of the feather star *Antedon bifida* (Echinodermata, Crinoidea). *J Anat* 183 (Pt 1):1-11.
- 1180 **Feder, H. M. 1959.** The food of the starfish, *Pisaster ochraceus* along the California coast. *Ecology* 40:721-724.
- Fein, A., and M. Terasaki. 2005.** Rapid increase in plasma membrane chloride permeability during wound resealing in starfish oocytes. *J Gen Physiol* 126:151-159.
- Ferguson, J. C. 1969.** Feeding, digestion and nutrition in Echinodermata. Pp. 71-100 in *Chemical Zoology*, I. M. F. a. B. T. S. (eds), ed. Academic Press, London.
- 1185 **Ferguson, J. C. 1990.** Hyperosmotic properties of the fluids of the perivisceral coelom and watervascular system of starfish kept under stable conditions. *Comp. Physiol. Biochem*:245-248.
- Ferguson, J. C., and C. W. Walker. 1991.** Cytology and function of the madreporite systems of the starfish *Henricia sanguinolenta* and *Asterias vulgaris*. *J Morphol* 210:1-11.

- 1190 **Feuda, R., and A. B. Smith. 2015.** Phylogenetic signal dissection identifies the root of starfishes. *PlosOne* **10**:14.
- FioRito, R., C. Leander, and B. Leander. 2016.** Characterization of three novel species of Labyrinthulomycota isolated from ochre sea stars (*Pisaster ochraceus*). *Marine Biology* **163**:10.
- Foret, T. W., and J. M. Lawrence 2001.** Variation in abundance of subcuticular bacteria in Florida echinoderms *Symbiosis* **31**:309-322.
- 1195 **Frakolaki, E., P. Kaimou, M. Moraiti, K. I. Kalliampakou, K. Karampetsou, E. Dotsika, P. Liakos, D. Vassilacopoulou, P. Mavromara, R. Bartenschlager et al. 2018.** The role of tissue oxygen tension in Dengue Virus replication. *Cells* **7**:22.
- Fredericks DN, & Relman DA. 1996.** Sequence-based identification of microbial pathogens: a reconsideration of Koch's postulates. *Clin. Micro. Rev.* **9**:18-33.
- 1200 **Frischer, M., R. F. Lee, A. R. Price, T. L. Walters, M. A. Bassette, R. Verdiyev, M. C. Torris, K. Bulski, P. J. Geer, S. A. Powell et al. 2017.** Causes, diagnostics, and distribution of an ongoing penaeid shrimp black gill epidemic in the U.S. South Atlantic Bight. *J. Shellfish Res* **36**:487-500.
- Fuess, L. E., M. E. Eisenlord, C. J. Closek, A. M. Tracy, R. Mauntz, S. Gignoux-Wolfsohn, M. M. Moritsch, R. Yoshioka, C. A. Burge, C. D. Harvell et al. 2015.** Up in arms: Immune and nervous
- 1205 system response to sea star wasting disease. *PLoS One* **10**:e0133053.
- Furukawa, R., Y. Takahashi, Y. Nakajima, M. Dan-Sohkawa, and H. Kaneko. 2009.** Defense system by mesenchyme cells in bipinnaria larvae of the starfish, *Asterina pectinifera*. *Dev Comp Immunol* **33**:205-215.
- Furukawa, R., K. Tamaki, H. Kaneko. 2016.** Two macrophage migration inhibitory factors regulate starfish larval immune cell chemotaxis. *Immunol Cell Biol* **94(4)**:315-21.
- 1210 **Gage, J. D., and P. A. Tyler. 1991.** *Deep-sea biology: a natural history of organisms at the deep-sea floor*. Cambridge University Press.

- Glockling, S. L., W. L. Marshall, and F. H. Gleason. 2013.** Phylogenetic interpretations and ecological potentials of the Mesomycetozoa (Ichthyosporea). *Fungal Ecology* **6**:237-247.
- 1215 **Goggin, C. L., and C. Bouland. 1997.** The ciliate *Orchitophrya* cf. *stellarum* and other parasites and commensals of the northern Pacific seastar *Asterias amurensis* from Japan. *Int J Parasitol* **27**:1415-1418.
- Gordon, S. 2016a.** Phagocytosis: An Immunobiologic Process. *Immunity* **44**:463-475.
- Gordon, S. 2016b.** Phagocytosis: The Legacy of Metchnikoff. *Cell* **166**:1065-1068.
- Gravem, S. A., W. N. Heady, V. R. Saccomanno, K. F. Alvstad, A. L. M. Gehman, T. N. Frierson, and S. L. Hamilton. 2021.** *Pycnopodia helianthoides*. *IUCN Red List of Threatened Species 2021*.
- 1220 **Gravem, S. A., and S. G. Morgan. 2017.** Shifts in intertidal zonation and refuge use by prey after mass mortalities of two predators. *Ecology* **98**:1006-1015.
- Gregg, A., M. Hatay, A. Haas, N. Robinett, K. Barott, M. Vermeij, K. Marhaver, P. Meirelles, F. Thompson, and F. Rohwer. 2013.** Biological oxygen demand optode analysis of coral reef-associated microbial communities exposed to algal exudates. *PeerJ* **1**:e107.
- 1225 **Groner, M. L., J. Maynard, R. Breyta, R. B. Carnegie, A. Dobson, C. S. Friedman, B. Froelich, M. Garren, F. M. Gulland, S. F. Heron et al. 2016.** Managing marine disease emergencies in an era of rapid change. *Philos Trans R Soc Lond B Biol Sci* **371**.
- Gudenkauf, B. M., J. B. Eaglesham, W. M. Aragundi, and I. Hewson. 2014.** Discovery of urchin-associated densovirus (family Parvoviridae) in coastal waters of the Big Island, Hawaii. *J Gen Virol* **95**:652-658.
- 1230 **Gudenkauf, B. M., and I. Hewson. 2015.** Metatranscriptomic Analysis of *Pycnopodia helianthoides* (Asteroidea) Affected by Sea Star Wasting Disease. *PLoS One* **10**:e0128150.
- Haas, A. F., C. E. Nelson, L. Wegley Kelly, C. A. Carlson, F. Rohwer, J. J. Leichter, A. Wyatt, and J. E. Smith. 2011.** Effects of coral reef benthic primary producers on dissolved organic carbon and microbial activity. *PLoS One* **6**:e27973.
- 1235 **Haggerty, J. M., and E. A. Dinsdale. 2016.** Distinct biogeographical patterns of marine bacterial taxonomy and functional genes. *Glob. Ecol. Biogeogr* **26**:177-190.

- Hall, M. R., K. M. Kocot, K. W. Baughman, S. L. Fernandez-Valverde, M. E. A. Gauthier, W. L.
1240 Hatleberg, A. Krishnan, C. McDougall, C. A. Motti, E. Shoguchi et al. 2017. The crown-of-thorns
starfish genome as a guide for biocontrol of this coral reef pest. *Nature* **544**:231-234.
- Hamel, J. F., S. Jobson, G. Caulier, and A. Mercier. 2021. Evidence of anticipatory immune and
hormonal responses to predation risk in an echinoderm. *Sci Rep* **11**:10691.
- Hardy, S. K., C. J. Deutsch, T. A. Cross, M. de Wit, and J. A. Hostetler. 2019. Cold-related Florida
1245 manatee mortality in relation to air and water temperatures. *PLoS One* **14**:e0225048.
- Harley, C., M. Pankey, J. Wares, R. Grosberg, and M. Wonham. 2006. Color polymorphism and
genetic structure in the sea star *Pisaster ochraceus*. *Biol Bull* **211**:248-262.
- Harrold, C., and J. S. Pearse. 1987. The ecological role of echinoderms in kelp forests. Pp. 37-233.
- Harvell, C. D., K. Kim, J. M. Burkholder, R. R. Colwell, P. R. Epstein, D. J. Grimes, E. E.
1250 Hofmann, E. K. Lipp, A. D. Osterhaus, R. M. Overstreet et al. 1999. Emerging marine diseases--
climate links and anthropogenic factors. *Science* **285**:1505-1510.
- Harvell, C. D., D. Montecino-Latorre, J. M. Caldwell, J. M. Burt, K. Bosley, A. Keller, S. F. Heron,
A. K. Salomon, L. Lee, O. Pontier et al. 2019. Disease epidemic and a marine heat wave are associated
with the continental-scale collapse of a pivotal predator (*Pycnopodia helianthoides*). *Science Advances*
1255 **5**:8.
- Haszprunar, G., C. Vogler, and G. Wörheide. 2017. Persistent Gaps of Knowledge for Naming and
Distinguishing Multiple Species of Crown-of-Thorns-Seastar in the *Acanthaster planci* Species Complex.
Diversity **9**:22.
- Held, M. B. E., and C. D. G. Harley. 2009. Responses to low salinity by the sea star *Pisaster ochraceus*
1260 from high- and low-salinity populations. *Invertebr. Biol.* **128**:381-390.
- Hendler, G., and D. R. Franz. 1982. The biology of a brooding seastar, *Leptasterias tenera*, in Block
Island Sound. *Biol. Bull.* **162**:273-289.

- Hewson, I. 2021.** Microbial respiration in the asteroid diffusive boundary layer influenced sea star wasting disease during the 2013–2014 northeast Pacific Ocean mass mortality event. *Marine Ecology Progress Series* in press.
- Hewson, I., C. A. Aquino, and C. M. DeRito. 2020a.** Virome Variation during Sea Star Wasting Disease Progression in *Pisaster ochraceus* (Asteroidea, Echinodermata). *Viruses* **12**:17.
- Hewson, I., K. S. I. Bistolas, E. M. Quijano Carde, J. B. Button, P. J. Foster, J. M. Flanzenbaum, J. Kocian, and C. K. Lewis. 2018.** Investigating the complex association between viral ecology, environment and Northeast Pacific Sea Star Wasting. *Frontiers in Marine Science* **5**:14.
- Hewson, I., J. B. Button, B. M. Gudenkauf, B. Miner, A. L. Newton, J. K. Gaydos, J. Wynne, C. L. Groves, G. Hendler, M. Murray et al. 2014.** Densovirus associated with sea-star wasting disease and mass mortality. *Proc Natl Acad Sci U S A* **111**:17278-17283.
- Hewson, I., M. R. Johnson, and I. R. Tibbetts. 2020b.** An Unconventional Flavivirus and Other RNA Viruses in the Sea Cucumber (Holothuroidea; Echinodermata) Virome. *Viruses* **12**.
- Hewson, I., and M. A. Sewell. 2021.** Surveillance of densoviruses and mesomycetozoans inhabiting grossly normal tissues of three Aotearoa New Zealand asteroid species. *PLoS One* **16**:e0241026.
- Hewson, I., B. Sullivan, E. W. Jackson, Q. Xu, H. Long, C. Lin, E. M. Quijano Cardé, J. Seymour, N. Siboni, M. R. L. Jones et al. 2019.** Perspective: Something old, something new? Review of wasting and other mortality in Asteroidea (Echinodermata). *Front. Mar. Sci* **6**:1-8.
- Hibino, T., M. Loza-Coll, C. Messier, A. J. Majeske, A. H. Cohen, D. P. Terwilliger, K. M. Buckley, V. Brockton, S. V. Nair, K. Berney et al. 2006.** The immune gene repertoire encoded in the purple sea urchin genome. *Dev Biol* **300**:349-365.
- Hildemann, W. H., and T. G. Dix. 1972.** Transplantation reactions of tropical Australian echinoderms. *Transplantation* **14**:624-633.
- Hoj, L., N. Levy, B. K. Baillie, P. L. Clode, R. C. Strohmaier, N. Siboni, N. S. Webster, S. Uthicke, and D. G. Bourne. 2018.** Crown-of-Thorns Sea Star *Acanthaster cf. solaris* Has Tissue-Characteristic Microbiomes with Potential Roles in Health and Reproduction. *Appl Environ Microbiol* **84**:18.

- 1290 **Holland, N. D., and K. H. Nealson. 1978.** Fine-structure of the echinoderm cuticle and the sub-cuticular
bacteria of echinoderms. *Acta Zoologica* **59**:169-185.
- Hughes, T. P. 1994.** Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef.
Science **265**:1547-1551.
- Hughes, T. P., K. D. Anderson, S. R. Connolly, S. F. Heron, J. T. Kerry, J. M. Lough, A. H. Baird,
1295 J. K. Baum, M. L. Berumen, T. C. Bridge et al. 2018a.** Spatial and temporal patterns of mass bleaching
of corals in the Anthropocene. *Science* **359**:80-83.
- Hughes, T. P., A. H. Baird, D. R. Bellwood, M. Card, S. R. Connolly, C. Folke, R. Grosberg, O.
Hoegh-Guldberg, J. B. Jackson, J. Kleypas et al. 2003.** Climate change, human impacts, and the
resilience of coral reefs. *Science* **301**:929-933.
- Hughes, T. P., J. T. Kerry, and T. Simpson. 2018b.** Large-scale bleaching of corals on the Great
1300 Barrier Reef. *Ecology* **99**:501.
- Hyman, L. 1955.** *The Invertebrates: Echinodermata*. New York ; London(etc) : McGraw-Hill.
- Iken, K., B. Konar, L. Benedetti-Cecchi, J. J. Cruz-Motta, A. Knowlton, G. Pohle, A. Mead, P.
Miloslavich, M. Wong, T. Trott et al. 2010.** Large-scale spatial distribution patterns of echinoderms in
nearshore rocky habitats. *PLoS One* **5**:1-14.
- 1305 **Ivanova, E. P., N. M. Gorshkova, T. Sawabe, N. V. Zhukova, K. Hayashi, V. V. Kurilenko, Y.
Alexeeva, V. Buljan, D. V. Nicolau, V. V. Mikhailov et al. 2004.** *Sulfitobacter delicatus* sp. nov. and
Sulfitobacter dubius sp. nov., respectively from a starfish (*Stellaster equestris*) and sea grass (*Zostera
marina*). *Int J Syst Evol Microbiol* **54**:475-480.
- Jackson, E. W., K. S. Bistolas, J. B. Button, and I. Hewson. 2016.** Novel Circular Single-Stranded
1310 DNA Viruses among an Asteroid, Echinoid and Holothurian (Phylum: Echinodermata). *PLoS One*
11:e0166093.
- Jackson, E. W., C. Pepe-Ranney, S. J. Debenport, D. H. Buckley, and I. Hewson. 2018.** The
Microbial Landscape of Sea Stars and the Anatomical and Interspecies Variability of Their Microbiome.
Front Microbiol **9**:1829.

- 1315 **Jackson, E. W., C. Pepe-Ranney, M. R. Johnson, D. L. Distel, and I. Hewson. 2020a.** A Highly Prevalent and Pervasive Densovirus Discovered among Sea Stars from the North American Atlantic Coast. *Appl Environ Microbiol* **86**:12.
- Jackson, E. W., R. C. Wilhelm, M. R. Johnson, H. L. Lutz, I. Danforth, J. K. Gaydos, M. W. Hart, and I. Hewson. 2020b.** Diversity of Sea Star-Associated Densoviruses and Transcribed Endogenous
- 1320 Viral Elements of Densovirus Origin. *J Virol* **95**.
- Jaffe, N., R. Eberl, J. Bucholz, and C. S. Cohen. 2019.** Sea star wasting disease demography and etiology in the brooding sea star *Leptasterias* spp. *PLoS One* **14**:e0225248.
- Jangoux, M. 1982.** *Food and feeding mechanisms*. A.A. Balkema, Rotterdam.
- Jensen, M. 1966.** The response of two sea-urchins to the sea-star *Marthasterias glacialis* (L.) and other
- 1325 stimuli. *Ophelia* **3**:209-219.
- Johnson, P. T. 1971.** Studies on diseased urchins from Point Loma. Annual Report Kelp Habitat Improvement Project. *California Institute of Technology, Pasadena*:82-90.
- Jurgens, L. J., L. Rogers-Bennett, P. T. Raimondi, L. M. Schiebelhut, M. N. Dawson, R. K Grosberg, and B. Gaylord. 2015.** Patterns of mass mortality among rocky shore invertebrates across
- 1330 100km of northeastern Pacific coastline. *PLoS One* **10**: e0131969.
- Karp, R. D., and W. H. Hildemann. 1976.** Specific allograft reactivity in the sea star *Dermasterias imbricata*. *Transplantation* **22**:434-439.
- Kay, S. W. C., A. M. Gehman, and C. D. G. Harley. 2019.** Reciprocal abundance shifts of the intertidal sea stars, *Evasterias troschelii* and *Pisaster ochraceus*, following sea star wasting disease. *Proc Biol Sci*
- 1335 **286**:20182766.
- Kelly, M. S., M. F. Barker, J. D. McKenzie, and J. Powell. 1995.** The Incidence and Morphology of Subcuticular Bacteria in the Echinoderm Fauna of New Zealand. *Biol Bull* **189**:91-105.
- Kerk, D., A. Gee, M. Standish, P. O. Wainright, and A. S. Drum. 1995.** The rosette agent of Chinook salmon (*Oncorhynchus tshawytscha*) is closely related to choanoflagellates, as determined by phylogenetic
- 1340 analyses of its small ribosomal subunit RNA. *Marine Biology* **122**:187-192.

- Kohl, W. T., T. I. McClure, and B. G. Miner. 2016.** Decreased Temperature Facilitates Short-Term Sea Star Wasting Disease Survival in the Keystone Intertidal Sea Star *Pisaster ochraceus*. *PLoS One* **11**:e0153670.
- Konar, B., T. J. Mitchell, K. Iken, H. Coletti, T. Dean, Esler, D. , M. Lindeberg, B. Pister, and B. Weitzman. 2019.** Wasting disease and static environmental variables drive sea star assemblages in the Northern Gulf of Alaska. *Journal of Experimental Marine Biology and Ecology* **520**:151-209.
- Lafferty, K., and T. Suchanek. 2016.** Revisiting Paine’s 1966 Sea Star Removal Experiment, the Most-Cited Empirical Article in the American Naturalist. *The American Naturalist* **188**.
- Landsberg, J. H., Y. Kiryu, E. C. Peters, P. W. Wilson, N. Perry, Y. Waters, K. E. Maxwell, L. K. Huebner, and T. M. Work. 2020.** Stony Coral Tissue Loss Disease in Florida Is Associated With Disruption of Host–Zooxanthellae Physiology. *Front. Mar. Sci.* **7**:24.
- Lauerman, L. M. L. 1998.** Diet and feeding behavior of the deep-water sea star *Rathbunaster californicus* (Fisher) in the Monterey Submarine Canyon. *Bull. Mar. Sci.* **63**:523-530.
- Lawrence , J. M. 2013.** *Starfish : biology and ecology of the Asteroidea*. Johns Hopkins University Press, Baltimore.
- Lawrence, J. M. 2015.** *Starfish: Biology and Ecology of the Asteroidea*, Vol. John Hopkins University Press, Washington DC.
- Leighton, B., J. D. G. Boom, C. Bouland, and M. J. Smith. 1991.** Castration and mortality in *Pisaster ochraceus* parasitized by *Orchitophrya stellarum* (Ciliophora). *Diseases of Aquatic Organisms* **10**:71-73.
- Lessios, H. A. 2016.** The Great *Diadema antillarum* Die-Off: 30 Years Later. *Ann Rev Mar Sci* **8**:267-283.
- Lessios, H. A., D. R. Robertson, and J. D. Cubit. 1984.** Spread of *Diadema* mass mortality through the Caribbean. *Science* **226**:335-337.
- Linchango Jr, G. V., D. W. Foltz, R. Reid, J. Williams, C. Nodzak, A. M. Kerr, A. K. Miller, R. Hunter, N. G. Wilson, W. J. Nielsen et al. 2017.** The phylogeny of extant starfish (Asteroidea:

- Echinodermata) including *Xyloplax*, based on comparative transcriptomics. *Molecular phylogenetics and evolution* **115**:161-170.
- Lloyd, M. M., and M. H. Pespeni. 2018.** Microbiome shifts with onset and progression of Sea Star Wasting Disease revealed through time course sampling. *Sci Rep* **8**:16476.
- 1370 **Luo, P., W. Wang, Y. Wang, C. Hu, and X. He. 2013.** *Pseudoalteromonas xishaensis* sp. nov., isolated from *Acanthaster planci* in the Xisha islands. *Antonie Van Leeuwenhoek* **104**:779-785.
- Mah, C. L., and D. B. Blake. 2012.** Global diversity and phylogeny of the Asteroidea (Echinodermata). *PloSOne* **7**:22.
- Majeske, A. J., C. J. Bayne, and L. C. Smith. 2013.** Aggregation of sea urchin phagocytes is augmented
1375 in vitro by lipopolysaccharide. *PLoS One* **8**:e61419.
- Martinez, A., A. C. Villaro, and P. Sesma. 1989.** Microscopic study of the pyloric caeca of the starfish *Marthasterias glacialis* (Echinodermata): Finding of endocrine cells. *J Morphol* **202**:151-164.
- Martinez, A. S., M. Byrne, and R. A. Coleman. 2016.** What and when to eat? Investigating the feeding habits of an intertidal herbivorous starfish. *Mar Biol* **163**:1-13.
- 1380 **Martinez, A. S., M. Byrne, and R. A. Coleman. 2017.** Filling in the grazing puzzle: A synthesis of herbivory in starfish. *Oceanography and Marine Biology: Annual Review* **55**:11-44.
- Matranga, V., R. Bonaventura, and G. Di Bella. 2002.** Hsp70 as a stress marker of sea urchin coelomocytes in short term cultures. *Cell Mol Biol (Noisy-le-grand)* **48**:345-349.
- McClintock, J. B., R. A. Angus, C. Ho, C. D. Amsler, and B. J. Baker. 2008.** A laboratory study of
1385 behavioral interactions of the Antarctic keystone sea star *Odontaster validus* with three sympatric predatory sea stars *Mar Biol* **154**:1077-1084.
- McClintock, J. B., and T. J. Robnett. 1986.** Size selective predation by the asteroid *Pisaster ochraceus* on the bivalve *Mytilus californianus*: A cost-benefit analysis. *Marine Ecology* **7**:321-332.
- McEdward, L. R., W. B. Jaeckle, and M. Komatsu. 2002.** *Phylum Echinodermata: Asteroidea*,
1390 Academic Press, London.

- McEdward, L. R., and D. A. Janies. 1993.** Life cycle evolution in asteroids: what is a larva? *Biological Bulletin* **184**:255-268.
- McEdward, L. R., and B. G. Miner. 2001.** Larval and life cycle patterns in echinoderms. *Canadian Journal of Zoology* **79**:1125-1170.
- 1395 **McKenzie, J. D., and I. V. Grigolava. 1996.** The echinoderm surface and its role in preventing microfouling. *Biofouling* **10**:261-272.
- McPherson, M. L., D. J. I. Finger, H. F. Houskeeper, T. W. Bell, M. H. Carr, L. Rogers-Bennett, and R. M. Kudela. 2021.** Large-scale shift in the structure of a kelp forest ecosystem co-occurs with an epizootic and marine heatwave. *Commun Biol* **4**:298.
- 1400 **Mead, A. D. 1898.** *Annual report of the Commissioners of Inland Fisheries made to the General Assembly / State of Rhode Island.*
- Melroy, L. M. and C. S. Cohen. 2021.** Temporal and spatial variation in population structure among brooding sea stars in the genus *Leptasterias*. *Ecol. Evol.* **11**: 3313-3331.
- Menge, B. 1979.** Coexistence between the seastars *Asterias vulgaris* and *A. forbesi* in a heterogenous environment: a non-equilibrium explanation. *Oecologia* **272**:245-272.
- 1405 **Menge, B., E. L. Berlow, C. A. Blanchette, S. A. Navarrete, and S. B. Yamada. 1994.** The Keystone Species Concept: Variation in Interaction Strength in a Rocky Intertidal Habitat. *Ecological Monographs* **64**:249-286.
- Menge, B. A., E. B. Cerny-Chipman, A. Johnson, J. Sullivan, S. Gravem, and F. Chan. 2016.** Sea Star Wasting Disease in the Keystone Predator *Pisaster ochraceus* in Oregon: Insights into Differential Population Impacts, Recovery, Predation Rate, and Temperature Effects from Long-Term Research. *PLoS One* **11**:e0153994.
- 1410 **Menge, J. L., and B. A. Menge. 1974.** Role of Resource Allocation, Aggression and Spatial Heterogeneity in Coexistence of Two Competing Intertidal Starfish. *Ecol. Monogr*:189-209.
- 1415 **Mercier, A., and J. F. Hamel. 2013.** *Reproduction in Asteroidea*. The Johns Hopkins Press, Baltimore MD USA.

- Miner, C. M., J. L. Burnaford, R. F. Ambrose, L. Antrim, H. Bohlmann, C. A. Blanchette, J. M. Engle, S. C. Fradkin, R. Gaddam, C. D. G. Harley et al. 2018.** Large-scale impacts of sea star wasting disease (SSWD) on intertidal sea stars and implications for recovery. *PLoS One* **13**:e0192870.
- 1420 **Miyoshi, K., Y. Kuwahara, and K. Miyashita. 2018.** Tracking the Northern Pacific sea star *Asterias amurensis* with acoustic transmitters in the scallop mariculture field of Hokkaido, Japan. *Fisheries science* **84**:349-355.
- Mladenov, P. V., and R. D. Burke. 1994.** *Echinodermata: asexual propagation*. Oxford and IBH Publications, New Delhi.
- 1425 **Mladenov, P. V., S. Igdoura, S. Asotra, and R. D. Burke. 1989.** Purification and partial characterization of an autotomy-promoting factor from the sea star *Pycnopodia helianthoides*. *Biol Bull* **176**:169-175.
- Montecino-Latorre, D., M. E. Eisenlord, M. Turner, R. Yoshioka, C. D. Harvell, C. V. Pattengill-Semmens, J. D. Nichols, and J. K. Gaydos. 2016.** Devastating Transboundary Impacts of Sea Star Wasting Disease on Subtidal Asteroids. *PLoS One* **11**:e0163190.
- 1430 **Mooi, R., and B. David. 2000.** What a new model of skeletal homologies tells us about asteroid evolution. *American Zoologist* **40**:326-339.
- Moritsch, M. M., and P. Raimondi. 2018.** Reduction and recovery of keystone predation pressure after disease-related mass mortality. *Ecology and Evolution* **8**:3952-3964.
- 1435 **Motokawa, T. 1982.** Rapid change in mechanical properties of echinoderm connective tissues caused by coelomic fluid. *Comp. Biochem. Physiol. Part C, Comp.* **73**:223-229.
- Motokawa, T. 1986.** Morphology of the spines and spine joint in the crown-of-thorns starfish *Acanthaster planci* (Echinodermata, Asteroidea). *Zoomorphology* **106**:247-253.
- Motokawa, T. 2011.** Mechanical mutability in connective tissue of starfish body wall. *Biol Bull* **221**:280-1440 289.
- Muehlstein, L. K., D. Porter, and F. T. Short. 1991.** *Labyrinthula zosterae* sp. nov., the Causative Agent of Wasting Disease of Eelgrass, *Zostera marina*. *Mycologia* **83**:180-191.

- Murie, K. A., and P. E. Bourdeau. 2019.** Predator identity dominates non-consumptive effects in a disease-impacted rocky shore food web. *Oecologia* **191**:945-956.
- 1445 **Myhre, S., and A. Acevedo-Gutiérrez. 2007.** Recovery of sea urchin *Diadema antillarum* populations is correlated to increased coral and reduced macroalgal cover. *Marine Ecology Progress Series* **329**:205-210.
- Nakagawa, S., H. Saito, A. Tame, M. Hirai, H. Yamaguchi, T. Sunata, M. Aida, H. Muto, S. Sawayama, and Y. Takaki. 2017.** Microbiota in the coelomic fluid of two common coastal starfish species and characterization of an abundant *Helicobacter*-related taxon. *Sci Rep* **7**:8764.
- 1450 **Narita, H., S. Matsubara, N. Miwa, S. Akahane, M. Murakami, T. Goto, M. Nara, T. Noguchi, T. Saito, Y. Shida et al. 1987.** *Vibrio-Alginolyticus*, a TTX producing bacterium isolated from the sea star *Astropecten polyacanthus*. *Bulletin of the Japanese Society of Scientific Fisheries* **53**:617-621.
- Nunez-Pons, L., T. M. Work, C. Angulo-Preckler, J. Moles, and C. Avila. 2018.** Exploring the pathology of an epidermal disease affecting a circum-Antarctic sea star. *Sci Rep* **8**:11353.
- 1455 **O'Neill, P. 1989.** Structure and mechanics of starfish body wall. *J Exp Biol* **147**:53-89.
- Ogawa, H., and E. Tanoue. 2003.** Dissolved Organic Matter in Oceanic Waters. *Journal of Oceanography* **59**:129-147.
- Olson, J. B., D. J. Gochfeld, and M. Slattery. 2006.** Aplysina red band syndrome: a new threat to Caribbean sponges. *Dis Aquat Organ* **71**:163-168.
- 1460 **Paine, R. T. 1966.** Food web complexity and species diversity. *Am Nat* **100**:65-75.
- Paine, V. L. 1926.** Adhesion of the tube feet in starfishes. *Journal of Experimental Zoology* **45**:361-366.
- Paps, J., L. A. Medina-Chacon, W. Marshall, H. Suga, and I. Ruiz-Trillo. 2013.** Molecular phylogeny of unikonts: new insights into the position of apusomonads and ancyromonads and the internal relationships of opisthokonts. *Protist* **164**:2-12.
- 1465 **Pearse, J. S. 1967.** Coelomic water volume control in the antarctic sea-star *Odontaster validus*. *Nature* **216**:1118-1119.

- Peters, E. C. 1992.** *Diseases of other invertebrate phyla: Porifera, Cnidaria, Ctenophora, Annelida, Echinodermata*, In: Couch JA (ed) Pathobiology of marine and estuarine organisms. CRC press, Gulf Breeze, Florida.
- 1470
- Petratis, P. S., and S. R. Dudgeon. 2020.** Declines over the last two decades of five intertidal invertebrate species in the western North Atlantic. *Commun Biol* **3**:591.
- Pincebourde, S., E. Sanford, and B. Helmuth. 2013.** Survival and arm abscission are linked to regional homeothermy in an intertidal sea star. *J Exp Biol* **216**:2183-2191.
- 1475
- Pinsino, A., and V. Matranga. 2015.** Sea urchin immune cells as sentinels of environmental stress. *Dev Comp Immunol* **49**:198-205.
- Pratchett, M. S., C. F. Caballes, J. C. Wilmes, S. Matthews, C. Mellin, H. P. A. Sweatman, L. E. Nadler, J. Brodie, C. A. Thompson, J. Hoey et al. 2017.** Thirty years of research on crown-of-thorns starfish (1986–2016): scientific advances and emerging opportunities. *Diversity* **9**:41.
- 1480
- Prusch, R. D., and F. Whoriskey. 1976.** Maintenance of fluid volume in the starfish water vascular system. *Nature* **262**:577-578.
- Ragan, M. A., C. L. Goggin, R. J. Cawthorn, L. Cerenius, A. V. Jamieson, S. M. Plourde, T. G. Rand, K. Soderhall, and R. R. Gutell. 1996.** A novel clade of protistan parasites near the animal-fungal divergence. *Proc Natl Acad Sci U S A* **93**:11907-11912.
- 1485
- Raimondi, P., R. Sagarin, R. Ambrose, C. Bell, M. George, S. Lee, D. Lohse, C. Miner, and S. Murray. 2007.** Consistent Frequency of Color Morphs in the Sea Star *Pisaster ochraceus* (Echinodermata:Asteriidae) across Open-Coast Habitats in the Northeastern Pacific. *Pacific Science* **61**:201-210.
- Ramsay, K., J. Turner, S. Vize, and C. Richardson. 2000.** A link between predator density and arm loss in the starfish *Marthasterias glacialis* and *Asterias rubens*. *J Mar Biol Assoc UK* **80**:565-566.
- 1490
- Rast, J. P., L. C. Smith, M. Loza-Coll, T. Hibino, and G. W. Litman. 2006.** Genomic insights into the immune system of the sea urchin. *Science* **314**:952-956.

- Rivera-Posada, J. A., M. Pratchett, A. Cano-Gomez, J. D. Arango-Gomez, and L. Owens. 2011a.** Injection of *Acanthaster planci* with thiosulfate-citrate-bile-sucrose agar (TCBS). I. Disease induction. *Dis Aquat Organ* **97**:85-94.
- 1495
- Rivera-Posada, J. A., M. Pratchett, A. Cano-Gomez, J. D. Arango-Gomez, and L. Owens. 2011b.** Refined identification of *Vibrio* bacterial flora from *Acanthaster planci* based on biochemical profiling and analysis of housekeeping genes. *Dis Aquat Organ* **96**:113-123.
- Rivera-Posada, J. A., M. Pratchett, and L. Owens. 2011c.** Injection of *Acanthaster planci* with
- 1500 thiosulfate-citrate-bile-sucrose agar (TCBS). II. Histopathological changes. *Dis Aquat Organ* **97**:95-102.
- Robilliard, G. A. 1971.** Feeding behaviour and prey capture in an asteroid, *Stylasterias forreri*. *Syesis* **4**:191-195.
- Robles, C. D., R. A. Desharnais, C. Garza, M. J. Donahue, and C. A. Martinez. 2009.** Complex equilibria in the maintenance of boundaries: experiments with mussel beds. *Ecology* **90**:985-995.
- 1505 **Rogers, T. L., H. K. Schultz, and J. K. Elliott. 2018.** Size-dependent interference competition between two sea star species demographically affected by wasting disease. *Marine Ecology Progress Series* **589**:167-177.
- Ruiz-Ramos, D. V., L. M. Schiebelhut, K. J. Hoff, J. P. Wares, and M. N. Dawson. 2020.** An initial comparative genomic autopsy of wasting disease in sea stars. *Mol Ecol* **29**:1087-1102.
- 1510 **Ruppert, E. E., R. S. Fox and R. D. Barnes. 2004.** *Echinodermata*. In: Invertebrate Zoology, Brooks/Cole-Thomson Learning, Belmont, CA.
- Santos, R., D. Haesaerts, M. Jangoux, and P. Flammang. 2005.** The tube feet of sea urchins and sea stars contain functionally different mutable collagenous tissues. *J Exp Biol* **208**:2277-2288.
- Schultz, J. A., R. N. Cloutier, and I. M. Cote. 2016.** Evidence for a trophic cascade on rocky reefs
- 1515 following sea star mass mortality in British Columbia. *PeerJ* **4**:e1980.
- Sea Urchin Genome Sequencing, C., E. Sodergren, G. M. Weinstock, E. H. Davidson, R. A. Cameron, R. A. Gibbs, R. C. Angerer, L. M. Angerer, M. I. Arnone, D. R. Burgess et al. 2006.** The genome of the sea urchin *Strongylocentrotus purpuratus*. *Science* **314**:941-952.

- Semmens, D. C., R. E. Dane, M. R. Pancholi, S. E. Slade, J. H. Scrivens, and M. R. Elphick. 2013.**
1520 Discovery of a novel neurophysin-associated neuropeptide that triggers cardiac stomach contraction and retraction in starfish. *J Exp Biol* **216**:4047-4053.
- Short, F. T., L. K. Muehlstein, and D. Porter. 1987.** Eelgrass Wasting Disease: Cause and Recurrence of a Marine Epidemic. *Biol Bull* **173**:557-562.
- Sieling, F. W. 1960.** Mass Mortality of the Starfish, *Asterias forbesi*, on the Atlantic Coast of Maryland.
1525 *Chesap. Sci* **1**:73-74.
- Silina, A. V. 2008.** Long-term changes in intra-and inter-specific relationships in a community of scallops and sea stars under bottom scallop mariculture. *Journal of Shellfish Research* **27**:1189-1194.
- Silva, J. R. 2000.** The onset of phagocytosis and identity in the embryo of *Lytechinus variegatus*. *Dev Comp Immunol* **24**:733-739.
- 1530 **Sloan, N. A. 1980.** Aspects of the feeding biology of asteroids. *Oceanogr. Mar. Biol. Anna. Rev.* **18** 57-124.
- Smith, J. E. 1937.** The structure and function of the tube feet in certain echinoderms. *Journal of the Marine Biological Association of the United Kingdom* **22**:345-357.
- Smith, L. C., V. Arizza, M. A. Barela Hudgell, G. Barone, A. G. Bodnar, K. M. Buckley, V. Cunsolo, N. M. Dheilly, N. Franchi, S. D. Fugmann et al. 2018.** *Echinodermata: The Complex Immune System in Echinoderms, in Advances in Comparative Immunology, E.L.*, Springer International Publishing: Cham.
- 1535 **Smith, L. C., J. Ghosh, K. M. Buckley, L. A. Clow, N. M. Dheilly, T. Haug, J. H. Henson, C. Li, C. M. Lun, A. J. Majeske et al. 2010.** Echinoderm immunity. *Adv Exp Med Biol* **708**:260-301.
- 1540 **Smith, V. J. 1981.** *The echinoderms, in Invertebrate blood cells*, Academic Press New York.
- Spalding, M. D., H. E. Fox, G. R. Allen, N. Davidson, Z. A. Ferdaña, M. Finlayson, B. S. Halpern, M. A. Jorge, A. Lombana, S. A. Lourie et al. 2007.** Marine Ecoregions of the World: A Bioregionalization of Coastal and Shelf Areas. *BioScience* **57**:573-583.

- 1545 **Stahli, A., R. Schaerer, K. Hoelzle, and G. Ribí. 2009.** Temperature induced disease in the starfish *Astropecten jonstoni*. *Mar. Biodivers. Rec* **2**:1-6.
- Stickle, W. B. , E. H. Weidner, and E. N. Kozloff. 2001.** Parasitism of *Leptasterias* spp. (Echinodermata: Asteroidea) by the ciliated protozoan *Orchitophrya stellarum* (Scuticociliata). *Invertebrate Biology* **120**:88-95.
- 1550 **Stickle, W. B., and G. J. Denoux. 1976.** Effects of *in situ* tidal salinity fluctuations on osmotic and ionic composition of body fluid in Southeastern Alaska Rocky intertidal fauna. *Mar. Biol.* **37**:125-135.
- Stickle, W. B., and E. N. Kozloff. 2008.** Association and distribution of the ciliate *Orchitophrya stellarum* with asteroiid sea stars on the west coast of North America. *Dis Aquat Organ* **80**:37-43.
- Strathmann, R. R. 1985.** Feeding and nonfeeding larval development and life history evolution in marine invertebrates. *Annual Review of Ecology and Systematics* **16**:339-361.
- 1555 **Sullivan, B. K., S. M. Trevathan-Tackett, S. Neuhauser, and L. L. Govers. 2018.** Review: Host-pathogen dynamics of seagrass diseases under future global change. *Mar Pollut Bull* **134**:75-88.
- Sunday, J., L. Raeburn, and M. W. Hart. 2008.** Emerging infectious disease in sea stars: castrating ciliate parasites in *Patiria miniata*. *Dis Aquat Organ* **81**:173-176.
- 1560 **Suzuki, G., S. Kai, and H. Yamashita. 2012.** Mass stranding of crown-of-thorns starfish. *Coral Reefs* **31**:821.
- Thorpe, J. P., and E. L. Spencer. 2000.** A mass stranding of the asteriod *Asterias rubens* on the Isle of Man. *J. Mar. Biol. Assoc. United Kingdom* **80**:749-750.
- Tiffany, W. J. 1978.** Mass mortality of *Luidia senegalensis* (Lamarck, 1816) on Captiva Island, Florida, with a note on its occurrence in Florida Gulf coastal waters. *Florida Sci.* **41**:63-64.
- 1565 **Tinoco, A. B., D. C. Semmens, E. C. Patching, E. F. Gunner, M. Egertova, and M. R. Elphick. 2018.** Characterization of NGFFYamide signaling in starfish reveals roles in regulation of feeding behavior and locomotory systems. *Front. Endocrinol* **9**:17.
- Tracy, A. M., M. L. Pielmeier, R. M. Yoshioka, S. F. Heron, and C. D. Harvell. 2019.** Increases and decreases in marine disease reports in an era of global change. *Proc Biol Sci* **286**:20191718.

- 1570 **Traiger, S., B. Konar, A. Doroff, and L. McCaslin. 2016.** Sea otters versus sea stars as major clam predators: evidence from foraging pits and shell litter. *Mar Ecol Prog Ser* **560**:73-86.
- Trotter, J. A., and T. J. Koob. 1989.** Collagen and proteoglycan in a sea urchin ligament with mutable mechanical properties. *Cell Tissue Res* **258**:527-539.
- Uthicke, S., B. Shaffelke, and M. Byrne. 2009.** A boom–bust phylum? Ecological and evolutionary
1575 consequences of density variations in echinoderms. *Ecological Monographs* **79**:3-24.
- Van Veldhuizen, H. D., and V. J. Oakes. 1981.** Behavioral responses of seven species of asteroids to the asteroid predator, *Solaster dawsoni*. *Oecologia* **48**:214-220.
- Van Volkom, K., L.G. Harris, and J. A Dijkstra. 2021.** Not all prey are created equal: Invasive ascidian diet mediates sea star wasting in *Henricia sanguinolenta*. *J. Exp. Mar. Biol. Ecol.* **544**: 151610.
- 1580 **Vevers, H. G. 1951.** The biology of *Asterias rubens* L. II. Parasitization of the gonads by the ciliate *Orchitophrya stellarum* Cope. *Journal of the Marine Biological Association of the United Kingdom* **29**:619-624.
- Vicknair, K., and J. A. Estes. 2012.** Interactions among sea otters, sea stars, and suspension-feeding invertebrates in the western Aleutian archipelago. *Mar Biol* **159**:2641-2649.
- 1585 **Wahlteitz, S. J., A. L. Newton, C. A. Harms, L. L. Lahner, and N. I. Stacy. 2020.** Coelomic fluid evaluation in *Pisaster ochraceus* affected by sea star wasting syndrome: Evidence of osmodyregulation, calcium homeostasis derangement, and coelomocyte responses. *Front Vet Sci* **7**:131.
- Wahlteitz, S. J., N. I. Stacy, L. L. Lahner, and A. L. Newton. 2019.** Coelomic fluid evaluation in clinically normal ochre sea stars *Pisaster ochraceus*: Cell counts, cytology, and biochemistry reference
1590 intervals. *J Aquat Anim Health* **31**:239-243.
- Wang, P, [Y. Chang](#), [J. Yu](#), [C. Li](#), [G. Xu](#). 2007.** Acute peristome edema disease in juvenile and adult sea cucumbers *Apostichopus japonicus* (Selenka) reared in North China. *J Invertebr Pathol* **96**(1):11-7.

- 1595 **Wares, J. P., and L. M. Schiebelhut. 2016.** What doesn't kill them makes them stronger: an association between elongation factor 1-alpha overdominance in the sea star *Pisaster ochraceus* and "sea star wasting disease". *PeerJ* **4**:e1876.
- Wilcox, M., and A. Jeffs. 2019.** Impacts of sea star predation on mussel bed restoration. *Restoration Ecology* **27(1)**:189-197.
- 1600 **Wilkie, I. C. 1979.** The juxtaligamental cells of *Ophiocomina nigra* (Abildgaard) (Echinodermata: Ophiuroidea) and their possible role in mechano-effector function of collagenous tissue. *Cell Tissue Res* **197**:515-530.
- Wilkie, I. C. 2002.** Is muscle involved in the mechanical adaptability of echinoderm mutable collagenous tissue? *J Exp Biol* **205**:159-165.
- 1605 **Wilkie, I. C., C. Carnevali, and F. Bonasoro. 1992.** The compass depressors of *Paracentrotus lividus* (Echinodermata, Echinoida): ultrastructural and mechanical aspects of their variable tensility and contractility. *Zoomorphology* **112**:143-153.
- Wobber, D. R. 1975.** Agonism in Asteroids. *Biol Bull* **148**:483-496.
- Work, T. M., T. M. Weatherby, C. M. DeRito, R. M. Besemer, and I. Hewson. 2021.** Sea Star Wasting Disease Pathology in *Pisaster Ochraceus* Shows a Basal-to-Surface Process Affecting Color Phenotypes Differently. *Diseases of Aquatic Organisms* **145**:21-33.
- 1610 **Zhang, Y., P. Tang, Y. Xu, W. Fang, X. Wang, Z. Fang, and Y. Xiao. 2016.** *Lutimaribacter marinistellae* sp. nov., isolated from a starfish. *Int J Syst Evol Microbiol* **66**:3675-3680.